Molecular Changes Induced in Rat Liver by Hemorrhage and Effects of Melanocortin Treatment

Caterina Lonati, Ph.D.,* Andrea Sordi, Ph.D.,* Daniela Giuliani, Ph.D.,† Luca Spaccapelo, M.D.,‡ Patrizia Leonardi, B.Sc.,§ Andrea Carlin, B.Sc.,§ Alessandra Ottani, Ph.D.,† Maria Galantucci, Ph.D.,# Paolo Grieco, Ph.D.,|| Anna Catania, M.D.,** Salvatore Guarini, Ph.D.††

ABSTRACT

Background: Melanocortin peptides improve hemodynamic parameters and prevent death during severe hemorrhagic shock. In the present research we determined influences of a synthetic melanocortin 1/4 receptor agonist on the molecular changes that occur in rat liver during hemorrhage.

Methods: Controlled-volume hemorrhage was performed in adult rats under general anesthesia by a stepwise blood withdrawal until mean arterial pressure fell to 40 mmHg. Then rats received either saline or the synthetic melanocortin 1/4 receptor agonist Butir-His-D-Phe-Arg-Trp-Sar-NH₂ (Ro27-3225; n = 6–8 per group). Hemogasanalysis was performed throughout a 60-min period. Gene expression in liver samples was determined at 1 or 3 h using quantitative real-time polymerase chain reaction.

Results: At 1 h, in saline-treated shocked rats, there were significant increases in activating transcription factor 3 (Atf3), early growth response 1 (Egr1), heme oxygenase (decycling) 1 (Hmox1), FB murine osteosarcoma viral oncogene homolog (Fos), and jun oncogene (Jun). These changes were prevented by Ro27-3225 treatment (A2m, 6.90 ± 3.44 vs. 2.37 ± 0.73, P < 0.001; Hspa1a, 10.34 ± 3.28 vs. 25.72 ± 3.64, P = 0.001; Hmox1, 3.28 ± 0.31 vs. 166.54 ± 35.03, P = 0.002; Jun, 6.62 ± 1.93 vs. 15.07 ± 2.09, P = 0.005; respectively). Increases in alpha-2-macroglobulin (A2m), heat shock 70kD protein 1A (Hspa1a), erythropoietin (Epo), and interleukin-6 (Il6) occurred at 3 h in shocked rats and were prevented by Ro27-3225 treatment (A2m, 6.90 ± 0.82 vs. 36.73 ± 4.00, P < 0.001; Hspa1a, 10.34 ± 3.28 vs. 25.72 ± 3.64, P = 0.001; Epo, 0.49 ± 0.13 vs. 2.37 ± 0.73, P = 0.002; Il6, 1.05 ± 0.15 vs. 1.88 ± 0.23, P < 0.001; respectively). Further, at 3 h in shocked rats treated with Ro27-3225 there were significant increases in...

What We Already Know about This Topic

• Melanocortin molecules, including adrenocorticotropic hormone, may protect organs against hemorrhagic shock-induced injury
• This study in rats investigates if a synthetic melanocortin may alleviate hemorrhagic-induced systemic and hepatic injury

What This Article Tells Us That Is New

• The melanocortin receptor agonist Ro27-3225 restores mean arterial pressure; attenuates metabolic acidosis; and inhibits upregulation of interleukin-6, acute phase protein A2m, and several other injury mediators in the liver after a hemorrhagic challenge

Copyright © 2012, the American Society of Anesthesiologists, Inc. Lippincott Williams & Wilkins. Anesthesiology 2012; 116:692–700

* Postdoctoral Fellow, Center for Surgical Research, ** Director, Center for Preclinical Investigation, Fondazione IRCCS Ca’ Granda - Ospedale Maggiore Policlinico, Milano, Italy. † Assistant Professor of Pharmacology, ‡ Postgraduate Student, § Postdoctoral Fellow, ¶ Full Professor of Pharmacology, Department of Biomedical Sciences, Section of Pharmacology, University of Modena and Reggio Emilia, Modena, Italy. # Research Assistant, Department of Internal Medicine, University of Milano, Milano, Italy. || Associate Professor of Pharmaceutical Chemistry, Department of Pharmaceutical Chemistry and Toxicology, University of Napoli “Federico II,” Napoli, Italy.

Received from the Department of Biomedical Sciences, Section of Pharmacology, University of Modena and Reggio Emilia, Modena, Italy. Submitted for publication August 5, 2011. Accepted for publication December 6, 2011. Supported by grant No. 20074Z8W3S from Fondazione Ca’ Granda Ospedale Maggiore Policlinico, Milano, Italy; and grant No. 050-08 from Fondazione Ca’ Granda Ospedale Maggiore Policlinico, Milano, Italy; and grant “Synthetic Melanocortins as a novel class of antiinflammatory drugs” from Fondazione Fiera, Milano, Italy.

Address correspondence to Dr. Guarini: Department of Biomedical Sciences, Section of Pharmacology, University of Modena and Reggio Emilia, Modena, Italy. salvatore.guarini@unimore.it. Information on purchasing reprints may be found at www.anesthesiology.org or on the masthead page at the beginning of this issue. ANESTHESIOLOGY’s articles are made freely accessible to all readers, for personal use only, 6 months from the cover date of the issue.

Supplemental digital content is available for this article. Direct URL citations appear in the printed text and are available in both the HTML and PDF versions of this article. Links to the digital files are provided in the HTML text of this article on the Journal’s Web site (www.anesthesiology.org).
EMORRHAGIC shock triggers a systemic inflammatory response and oxidative stress. Blood loss, oxidative stress, and the adrenergic outflow start signaling events that promote induction of genes involved in stress and heat shock response, inflammatory reaction, and apoptosis. These processes can cause systemic inflammation and organ damage. In the most severe cases, multiple organ dysfunction syndrome occurs as a serious complication associated with high morbidity and mortality.\(^1\)\(^{–}\)\(^8\) Redistribution of cardiac output and persistent gut ischemia after apparently adequate resuscitation can lead to hemorrhagic shock.

Treatment of hemorrhagic shock includes rapid operative resuscitation with fluids, blood transfusions, and vasopressor agents to limit activation of the inflammation mediator systems and abort the microcirculatory changes.\(^9\) However, these approaches may not be sufficient to prevent organ failure. Therefore, therapies against this pathologic condition should modulate multiple cellular events induced by hemorrhage.

Melanocortin peptides, the collective name for adrenocorticotropic hormone and α-, β-, and γ-melanocyte stimulating hormone, exert protective influences on the host during local and systemic injury.\(^10\) Effects of melanocortins are mediated by activation of five melanocortin (MC\(_1\) through MC\(_6\)) receptors that belong to the class A of G-protein-coupled seven transmembrane receptors.\(^10\) The melanocortin 4 (MC\(_4\)) is the prevalent melanocortin receptor within the central nervous system, where it is highly expressed in the hypothalamus, spinal cord, vagus nuclei, and cortex.\(^13\) The MC\(_1\) subtype is expressed in a wide range of peripheral tissues,\(^12\) including the human liver.\(^13\)

In previous research, we found that melanocortin peptides have a life-saving activity in experimental and clinical hemorrhagic shock.\(^14\)\(^{–}\)\(^18\) Indeed, early treatment with adrenocorticotropic hormone-(1–24) in a rat model of hemorrhagic shock prolonged survival and extended by some hours the time-limit for blood reinfusion to be effective.\(^17\) The melanocortin-induced reversal of hemorrhagic shock was associated with mobilization of the residual blood, that, in shock conditions, is trapped in capillaries and large blood reservoirs, including the liver and spleen.\(^19\) Furthermore, the survival rate in patients with acute type-A aortic dissection treated with adrenocorticotropic hormone-(1–24) was significantly increased.\(^18\) The synthetic MC\(_1\)/MC\(_4\) receptor agonist Ro27-3225 prevented death in rats subjected to lethal hemorrhagic shock.\(^14\) Such life-saving effect was associated with improved cardiovascular and respiratory functions, reduced concentration of plasma free radicals, and prevention of multiple organ damage. These results prompted further investigations on molecular effects of this molecule in hemorrhagic shock. Research was focused on the liver, which is highly susceptible to ischemic damage,\(^20\) and inflammatory cells accumulated in the liver during shock should be sources of inflammatory mediators that greatly contribute to multiple organ dysfunction.\(^14\)\(^{–}\)\(^16\)

The aim of the present study was to determine effects of melanocortin treatment on gene expression profile in the liver during hemorrhage. Transcript selection was made to examine class of representative molecules that could be significant in local and systemic injury after hemorrhage. A controlled volume nonlethal hemorrhage was used in order to perform prolonged analysis of gene expression. Ro27-3225 was chosen because of its selective affinity for the MC\(_1\) and MC\(_4\) receptor subtypes. Indeed, these two receptors mediate most of peripheral (MC\(_1\)) and brain-mediated (MC\(_4\)) antiinflammatory effects of melanocortins.\(^10\)

### Materials and Methods

#### Animals

Wistar rats (Harlan, Milan, Italy) were kept in air-conditioned colony rooms (temperature 21 ± 1°C; humidity 60%) on a natural light/dark cycle, with food in pellets and tap water available ad libitum. Body weight (270–300 g) was similar in all groups (\(n = 6–8\) per group; \(P > 0.05\)). Housing conditions and experimental procedures were in strict accordance with the European Community regulations on the use and care of animals for scientific purpose (CEE Council 89/609; Italian D.L.22-1-92 No. 116), and were approved by the Committee on Animal Health and Care of Modena and Reggio Emilia University (Comitato Etico per

---

**Fig. 1.** Treatment with the melanocortin MC\(_1\)/MC\(_4\) receptor agonist Ro27-3225 significantly restored mean arterial pressure values in rats subjected to controlled-volume hemorrhage. Values are means ± SEM (\(n = 6\)). *\(P < 0.01\) versus hemorrhage + saline. MAP = mean arterial pressure.
Table 1. Effect of Intravenous Injection of Ro27-3225 (90 μg/kg) on Venous pH, P O2, P CO2, HCO3−, SBE, SO2, and Lactate Levels in Hemorrhage Shocked Rats

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Time of Sampling</th>
<th>Rats (n)</th>
<th>pH</th>
<th>P O2 (mmHg)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Saline</td>
<td>Before bleeding</td>
<td>8</td>
<td>7.32 ± 0.03</td>
<td>54.20 ± 3.64</td>
</tr>
<tr>
<td></td>
<td>After bleeding</td>
<td>7</td>
<td>7.21 ± 0.01*</td>
<td>26.09 ± 0.81*</td>
</tr>
<tr>
<td></td>
<td>15 min after treatment</td>
<td>8</td>
<td>7.15 ± 0.03*</td>
<td>22.64 ± 0.51*</td>
</tr>
<tr>
<td></td>
<td>60 min after treatment</td>
<td>6</td>
<td>7.12 ± 0.02*</td>
<td>23.56 ± 0.53*</td>
</tr>
<tr>
<td>Ro27–3225</td>
<td>Before bleeding</td>
<td>8</td>
<td>7.30 ± 0.02</td>
<td>53.16 ± 2.98</td>
</tr>
<tr>
<td></td>
<td>After bleeding</td>
<td>8</td>
<td>7.18 ± 0.02*</td>
<td>25.69 ± 0.78*</td>
</tr>
<tr>
<td></td>
<td>15 min after treatment</td>
<td>7</td>
<td>7.14 ± 0.02*</td>
<td>38.02 ± 0.24†</td>
</tr>
<tr>
<td></td>
<td>60 min after treatment</td>
<td>8</td>
<td>7.28 ± 0.01†</td>
<td>45.39 ± 1.28†</td>
</tr>
</tbody>
</table>

Values are means ± SEM.
* P < 0.05, versus the corresponding value before bleeding. † P < 0.001, versus the corresponding value after bleeding.
HCO3− = bicarbonate; P CO2 = carbon dioxide partial pressure; P O2 = oxygen partial pressure; SBE = standard base excess; SO2 = oxygen saturation values.

Surgical Procedures, Shock Induction, and Treatments

Surgical procedures were performed under general anesthesia (urethane, 1.25 g/kg intraperitoneally). Urethane was chosen because it provides long-lasting and stable anesthesia with minimal interference with neurally mediated cardiovascular and respiratory regulatory functions.14–17,21 Animals were spontaneously ventilating. After heparinization (heparin sodium, 600 U/kg intravenously), rats were instrumented with indwelling polyethylene catheters in a common carotid artery to record arterial blood pressure and in iliac vein for treatments and bleeding. The arterial catheter was connected to a pressure transducer coupled to a polygraph (Mortara-Rangoni, Bologna, Italy). Hemorrhage was induced by a stepwise withdrawal of about 40% of circulating blood (1.4–1.6 ml/100 g body weight; n = 6–8 per group; P > 0.05), until mean arterial pressure (MAP) fell to and stabilized at 40 mmHg or fewer. The procedure was completed within 20 min. Sham hemorrhage rats were subjected to all surgical procedures of hemorrhage animals, but were not bled. All the procedures were performed in sterile conditions. Animals were randomly assigned to the following intravenous bolus treatments: 1) the MC1/MC5 receptor agonist Butir-His-D-Phe-Arg-Trp-Sar-NH2 (Ro27-3225) administered to hemorrhage rats, at the dose of 90 μg/kg dissolved in a volume of 1 ml/kg saline, 5 min after termination of the bleeding procedure; 2) hemorrhage and 3) sham hemorrhage rats received as control injection an equal volume of saline (1 ml/kg); 4) a further group of sham hemorrhage animals received Ro27-3225 (90 μg/kg dissolved in 1 ml/kg saline). Ro27-3225 was synthesized in our laboratory by conventional solid phase chemistry, purified by reversed phase high-performance liquid chromatography, and checked for proper molecular weight by mass-spectroscopy, as previously reported.14 Ro27-3225 was used at a fixed dose previously found to be effective in reversing severe hemorrhagic shock in rats in our laboratory.14

For gene expression study, rats treated with Ro27-3225 dissolved in saline (group 1: hemorrhage + Ro27-3225), or control rats treated with saline alone (group 2: hemorrhage + saline; and group 3: sham hemorrhage + saline), were sacrificed under deep anesthesia at 1 h (n = 6 per group) or 3 h (n = 6 per group) after treatment. Six normal (naïve) rats were sacrificed under deep anesthesia at time 0 and received no injections (baseline control for gene expression study). In all assessments, each animal was used for a single sample. Livers were immediately harvested and tissue samples were snap-frozen in liquid nitrogen for RNA extraction. Arterial blood pressure was monitored in all animals until sacrifice at scheduled intervals. The MAP values over time were compared among experimental groups 1–4 considering only animals that were sacrificed at 3 h (n = 6 per group).

Hemogasanalysis and Lactic Acid Measurement

Hemogasanalysis and lactic acid measurements were performed in experimental groups 1–4, in 6–8 rats per group and per time point. The recorded values over time of group 1 (hemorrhage + Ro273225) and group 2 (hemorrhage + saline) were compared. In all assessments, each animal was used for a single sample to avoid possible alterations of parameters that were recorded, induced by repeated blood sampling. Blood samples (0.2 ml) were taken from the venous catheter just before bleeding (basal conditions), immediately before treatment, and after 15 and 60 min after treatment, and analyzed for pH, oxygen partial pressure, carbon dioxide partial pressure, bicarbonate, standard base excess, and oxygen saturation by using a System 1302 pH/blood gas analyzer (Instrumentation Laboratory, Milan, Italy). Lactate was measured by means of an enzymatic test (Ortho Clinical Diagnostics, Rochester, NY).

RNA Isolation and Real-time Polymerase Chain Reaction

Gene expression was evaluated as described previously.22 In brief, total RNA was isolated on an ABI Prism 6100 Nucleic
Acid PrepStation using Total RNA Chemistry (Applied Biosystems, Carlsbad, CA). RNA was checked for integrity by electrophoresis on denaturing agarose-formaldehyde gels and quantified by optical density measurement at 260 nm. Real-time polymerase chain reaction analysis was based on TaqMan chemistry. Two micrograms of total RNA were reverse transcribed to single-stranded complementary DNA using the High-Capacity Complementary DNA Archive Kit (Applied Biosystems). Polymerase chain reactions were performed on an ABI Prism 7900HT Sequence Detection System (Applied Biosystems). Assay IDs for target and reference genes are reported in Supplemental Digital Content 1, which is a table listing all the genes investigated in this study. Fluorescence intensities were converted in threshold cycles (Ct) using reverse-transcribed Rat Universal Reference Total RNA (Clontech, Palo Alto, CA) as calibrator sample. Polymerase chain reaction efficiencies (η) were evaluated by reference complementary DNA standard curves covering a 4 to 5 log dynamic range. Relative quantities were normalized using a normalization factor calculated by the public algorithm geNorm VBA applet version 3.4 for Excel. Three independent polymerase chain reaction amplification experiments were performed for each transcript. The relative expression was calculated as the mean of normalized quantities in the replicates.

### Data Analysis and Statistical Evaluation

Analysis was performed blind to the treatment. Agglomerative hierarchical cluster analysis was performed on relative quantities of target genes using DNA-chip analyzer software.‡ In the analysis of genes listed in Supplemental Digital Content 1 (see http://links.lww.com/ALN/A812), which play a role in hemorrhagic shock, Spearman rank correlation and average linkage were used as similarity metric and clustering technique, respectively. Here, Spearman rank correlation and average linkage don’t test for significance the results; instead, they are used to show a global trend of gene expression across all the samples. Briefly, Spearman rank correlation, a nonparametric similarity measure, clusters genes whose expression levels may be very different, and it is robust against outliers. The average linkage method, used as distance metric, computes the distance between two gene clusters as the average of the distances between all the points in those clusters. Statistical evaluation of gene expression was performed on selected genes considered of relevant interest in our experimental conditions, on biologic basis.

Comparison of MAP values was performed using two-way repeated measures ANOVA and post hoc Bonferroni correction for comparisons of means. Differences in hemogasanalytic data, lactate levels, body weight, and blood withdrawn to induce shock were analyzed using one-way ANOVA followed by Bonferroni multiple comparison test. Differences in relative expression of individual transcripts were analyzed using two-way ANOVA followed by Bonferroni multiple comparison test. We assessed whether the different experimental groups differ among them in terms of the parameters we considered, such as MAP, gene expression, hemogasanalytic parameters, and lactate levels. We, therefore, set α (level of statistical significance) equal to 0.05 (two-tailed). Results with P < α were considered as statistically significant. All the statistical tests were performed using SigmaStat software, version 3.5 (Systat Software Inc, San Jose, CA).

### Results

MAP values after hemorrhage and the effects of Ro27-3225 treatment on such values are reported in figure 1. Ro27-3225 (n = 6) significantly enhanced MAP values relative to untreated animals (n = 6) during the whole observation period. Bleeding caused a significant alteration in pH, oxygen partial

### Table 1. Continued

<table>
<thead>
<tr>
<th>PCO₂ (mmHg)</th>
<th>HCO₃⁻ (mequiv/l)</th>
<th>SBE (mequiv/l)</th>
<th>SO₂ (%)</th>
<th>Lactate (mmol/l)</th>
</tr>
</thead>
<tbody>
<tr>
<td>46.01 ± 1.64</td>
<td>22.45 ± 0.62</td>
<td>-4.02 ± 0.49</td>
<td>84.01 ± 1.19</td>
<td>5.85 ± 0.51</td>
</tr>
<tr>
<td>52.39 ± 1.25*</td>
<td>19.01 ± 0.96*</td>
<td>-8.05 ± 0.72*</td>
<td>30.26 ± 1.18*</td>
<td>10.12 ± 1.01*</td>
</tr>
<tr>
<td>56.69 ± 1.95*</td>
<td>16.64 ± 1.12*</td>
<td>-12.24 ± 1.12*</td>
<td>19.64 ± 1.24*</td>
<td>11.01 ± 1.31*</td>
</tr>
<tr>
<td>58.32 ± 2.01*</td>
<td>15.91 ± 1.16*</td>
<td>-15.38 ± 1.32*</td>
<td>16.12 ± 0.84*</td>
<td>11.89 ± 1.28*</td>
</tr>
<tr>
<td>48.64 ± 1.31</td>
<td>24.01 ± 0.26</td>
<td>-3.86 ± 0.38</td>
<td>82.19 ± 1.91</td>
<td>5.69 ± 0.62</td>
</tr>
<tr>
<td>53.84 ± 1.61*</td>
<td>19.89 ± 0.62*</td>
<td>-8.09 ± 0.84*</td>
<td>29.18 ± 1.68*</td>
<td>10.31 ± 1.05*</td>
</tr>
<tr>
<td>64.32 ± 1.98*</td>
<td>18.84 ± 0.29*</td>
<td>-10.96 ± 0.49*</td>
<td>51.32 ± 1.28*†</td>
<td>10.84 ± 1.26*</td>
</tr>
<tr>
<td>47.39 ± 2.03</td>
<td>22.87 ± 0.98</td>
<td>-4.19 ± 0.83†</td>
<td>72.84 ± 3.64†</td>
<td>6.02 ± 0.58†</td>
</tr>
</tbody>
</table>
pressure, carbon dioxide partial pressure, bicarbonate, standard base excess, and oxygen saturation values, as well as in lactate levels (n = 6 – 8; table 1); treatment with Ro27-3225 almost completely restored the basal conditions throughout the 60-min observation period (n = 6 – 8; table 1). Treatment of sham animals with intravenous injection of Ro27-3225 or a control injection of an equal volume of saline (1 ml/kg) did not affect MAP (n = 6 per group; fig. 1), hemogasanalysis parameters, and lactate levels (n = 6 per group; not shown). Gene expression in the liver during hemorrhage was considerably altered (fig. 2). Indeed, out of the 46 transcripts examined, expression of nine genes was significantly different in saline-treated shocked animals relative to sham rats at either 1 or 3 h. At 1 h, in saline-treated hemorrhage animals, there were significant increases in activating transcription factor 3 (Atf3), early growth response 1 (Egr1), heme oxygenase (decycling) 1 (Hmox1), FBJ murine osteosarcoma viral oncogene homolog, and jun oncogene at 1 h; expression of alpha-2-macroglobulin, heat shock 70kD protein 1A, erythropoietin, and interleukin-6 was significantly reduced by treatment at 3 h. Bars denote mean ± SEM (n = 6). ** P < 0.01, *** P < 0.001. A2m = alpha-2-macroglobulin; Atf3 = activating transcription factor 3; Egr1 = early growth response 1; Epo = erythropoietin; Fos = FBJ murine osteosarcoma viral oncogene homolog; Hmox1 = heme oxygenase (decycling) 1; Hspa1a = heat shock 70kD protein 1A; Ile6 = interleukin-6; Jun = jun oncogene.

**Discussion**
The present research on hemorrhagic shock shows that administration of the MC1/MC4 receptor agonist Ro27-3225...
Liver biology is highly susceptible to ischemic injury associated with hemorrhagic shock and impairment in liver function widely affects other organs. In view of the consequences of hemorrhage-related liver injury, the present study investigated expression of representative genes whose induction seemingly triggers crucial signaling pathways during hemorrhage. The data showed marked up-regulation of genes related to immediate/early response (Jun, Fos, Atp3), acute phase response (A2m, Fga), response to oxidative stress (Hmox1, Epo, endothelin1 [Edn1], Hspa1a), and inflammation (Egr-1, II-6, s100 calcium binding protein A8, calgranulin A [s100a8], s100 calcium binding protein A9, calgranulin B [s100a9], chemokine [C-C motif] ligand 2 [Ccl2]). Treatment with the MC1/MC4 agonist Ro27-3225 was associated with impressive stability in gene expression. Indeed, several genes that were enhanced by hemorrhage showed only marginal changes when animals were treated with Ro27-3225. Gene profile in these livers was remarkably similar to the profile of organs from sham-shocked rats. It appears, therefore, that MC1/MC4 stimulation can prevent inflammatory responses induced by hemorrhage. This action could be very important to reduce subsequent organ dysfunction.

Melanocortin treatment prevented induction of key transcripts related to acute phase reaction and inflammatory response. The cytokine IL-6 promotes induction of acute phase proteins and its importance in posthemorrhage inflammation and organ damage/dysfunction is well established. Therefore, the observation that Ro27-3225 prevented IL-6 induction in the liver is particularly significant. The acute phase protein A2m was likewise raised by hemorrhage and inhibited by Ro27-3225. This observation reinforces the idea that hepatic acute phase reaction, a critical event in hemorrhage-induced systemic inflammatory response, is modulated by melanocortin treatment. In the present research, we found significant up-regulation of Egr-1 in hemorrhage rats; such increase was prevented by Ro27-3225 administration. Based on its multiple effects, the control of Egr-1 activation could be critical to protect liver tissue. This gene is induced by a variety of acute cellular stresses and is a transcription factor for several key mediators of inflammation, coagulation, vascular permeability, and injury.

Consistently, recent research found that Egr-1 messenger RNA was rapidly up-regulated in the liver during hemorrhagic shock, where it promoted local and systemic inflammatory reactions. Of interest, deficiency in Egr-1 resulted in a blunted inflammatory response, thus providing evidence that Egr-1 is an inducer of the early inflammatory response to shock that subsequently leads to systemic injury.

In evaluation of the effects of Ro27-3225 on gene expression in the hemorrhage liver, it should be considered that melanocortins participate in central regulation of cardiovascular functions. For example, α-melanocyte-stimulating hormone acutely increased blood pressure and heart rate through central stimulation of sympathetic nervous outflow. This action of the peptide was mediated by stimulation of MC4 receptors within the brain. Other research found that intracerebroventricular injections of α-melanocyte-stimulating hormone increased arterial pressure and renal sympathetic nerve activity in rabbits. However, in our previous investigations and in the present study, nanomolar concentrations of melanocortins did not affect MAP in sham (unshocked) animals.

Liver protection was not restricted to prevention of inflammation. In livers from hemorrhage rats treated with Ro27-3225 there was no up-regulation of immediate-early genes (Jun, Fos) and stress-induced genes (Atp3, Hmox1, and Hspa1a) that were conversely enhanced in untreated animals. The up-regulation of these transcripts in untreated...
hormone levels. Therefore, in shock conditions melanocortin-based treatment could be helpful drugs for a safe, non-toxic treatment able to rapidly improve cardiovascular function and tissue perfusion for some hours, as previously reported.17,18 By blocking the main pathophysiological mechanisms of organ damage, melanocortins would considerably extend the effective time-limit for blood reinfusion.17,18 The antishock effect of melanocortins in experimental and clinical conditions have been recently highlighted by Corander et al. in an authoritative review aimed at spreading these relevant findings of basic science to clinicians.

Some weakness of the present results for a potential clinical use of melanocortins in shock conditions deserve consideration. The animal model used in this research is closer to spontaneous bleeding in humans, and urethane, used as anesthetic, cannot be employed in the clinical setting. Therefore, the actual possibility to translate the present results to trauma patients in hemorrhagic shock conditions should be
verified in further animal models. Furthermore, the melanocortin-induced improvement in circulatory condition is transient, and requires volume restoration within a few hours. Conversely, the short half-life of peptide molecules, including natural melanocortins, that are broken down readily in the body fluids, should not represent a limitation as several of the generated fragments are biologically active. Finally, passage through the blood-brain barrier and lack of receptor selectivity of melanocortins should not be problematic in short-term, acute treatments such as in hemorrhagic shock.

References
survival in experimental hemorrhagic shock. Eur J Pharmacol 1986; 130:19–26

33. Maxwell MA, Muscat GE: The NR4A subgroup: immediate
early response genes with pleiotropic physiological roles.
Nucl Recept Signal 2006; 4:e002

34. Song KH: Orphan nuclear receptor Nur77 participates in

35. Pei L, Waki H, Vaitheesvaran B, Wilpitz DC, Kurland JJ,

36. Torres Filho IP, Torres LN, Pittman RN: Early physiologic
responses to hemorrhagic hypotension. Transl Res 2010;
155:78–88

37. Torres LN, Torres Filho IP, Barbee RW, Tiba MH, Ward KR,
Pittman RN: Systemic responses to prolonged hemorrhagic
hypotension. Am J Physiol Heart Circ Physiol 2004; 286:
H1811–20

38. Harhaj NS, Antonetti DA: Regulation of tight junctions and
loss of barrier function in pathophysiology. Int J Biochem

39. Blum MS, Toninelli E, Anderson JM, Balda MS, Zhou J,
O’Donnell L, Pardi R, Bender JR: Cytoskeletal rearrangement

RL, Fink MP: IL-6 is essential for development of gut barrier
dysfunction after hemorrhagic shock and resuscitation in
mice. Am J Physiol Gastrointest Liver Physiol 2003; 285:
G621–9

41. Fink MP, Delude RL: Epithelial barrier dysfunction: A unifying
theme to explain the pathogenesis of multiple organ dysfunction

42. Ichiyama T, Sakai T, Catania A, Barsh GS, Furukawa S, Lipton
JM: Inhibition of peripheral NF-κB activation by central ac-
tion of α-melanocyte-stimulating hormone. J Neuroimmunol
1999; 99:211–7

43. Macaluso A, McCoy D, Ceriani G, Watanabe T, Biltz J, Catania
A, Lipton JM: Antiinflammatory influences of α-MSH mole-
cules: Central neurogenic and peripheral actions. J Neurosci
1994; 14:2377–82

44. Giuliani D, Ottani A, Altavilla D, Bazzani C, Squadrito F,
Guarini S: Melanocortins and the cholinergic anti-inflamma-

45. Corander MP, Fenech M, Coll AP: Science of self-preservation:
How melanocortin action in the brain modulates body
weight, blood pressure, and ischemic damage. Circulation
2009; 120:2260–8