To the Editor:—We read with interest the correspondence by Zimmer et al.1 Although we agree with their conclusion that human error related to magnetic resonance imaging (MRI) use can only be minimized by adequate training, we believe lessening the risks of MRI technology itself is of equal importance. We feel it is time to stress the importance of “anesthesia-compatible” MRI, rather than putting all the emphasis on anesthesiologists adapting to the needs of the MRI machine. Anesthesiologists and patients are now forced into working under conditions that are far less than optimal in MRI suites that are cold and dark, have noisy equipment and facilities, and are often located far away from the main operating area.

There are three aspects of MRI that are important to the anesthesiologist: 1) avoidance of materials and equipment that will be attracted to the MRI machine, 2) avoidance of anesthetic devices that interfere with the function of the MRI machine, and 3) avoidance of MRI interference with the patient and the functioning of materials and devices used for anesthesia. This third aspect is often neglected: most MRI machines are not “anesthesia-compatible.”

The first point is obvious, well known, and scary but can be handled with a little experience.2 It is, however, extremely important for everyone to realize that complete elimination of the use of ferromagnetic materials in devices used in MRI suites is not feasible and is sometimes impossible, as was made clear in the letter by Zimmer et al.,1 among others.3 We have successfully dealt with this problem by anchoring all devices that have ferrous materials in them to a movable ceiling pendant system with a predetermined limited range of movement. Installation of metal detectors (similar to those used in airports) at the entrance to MRI suites can help to some degree.

The second point involves the use of equipment such as ventilators and infusion pumps for treatment and various patient monitors, personal computer-related devices, and local area network connections for medical information. MRI technology now frequently forces the anesthesiologist to discontinue the use of these devices during MRI activity despite their importance for patient safety.4 Alternative “MRI-compatible” devices are not always available, functional, or suited for critically ill patients, causing anesthesiologists to make an uncomfortable choice between the continuity of safety of treatment and MRI diagnosis.

The third aspect is the most difficult to resolve. We think more attention should be focused on alleviating this problem although some MRI properties, such as magnetic attraction, electric shock, or heating as a result of radiofrequency pulsing, seem to be inevitable.4 Anesthesiologists have been forced to adapt to MRI technology, raising a never-ending list of incompatibility issues. While working to build a new MRI suite, we realized that although there were few technical difficulties to overcome, lack of awareness of the issues involved with traditional MRI was playing a key role in holding back the development of more patient-friendly MRI technology. Companies attempted to work with that already make both MRI and anesthesiology-related equipment did not seem to find safety for patients under anesthesia during MRI a compelling enough reason to consider revising their MRI devices. It would be much more cost effective and safe to improve MRI machines and their installation, including the architectural design of MRI suites, than it would be to carry out patchwork renovation of numerous patient care devices.

The demand for anesthesia care in MRI suites continues to increase as interventional procedures using MRI continue to increase in frequency.5–7 Time spent in MRI suites will only become longer. Anesthesiologists, as advocates for patients, should actively voice their concern to improve MRI technology not only in terms of radiologic diagnostic function but also in terms of working environment, duration of examination, and, most importantly, patient safety. We must seek solutions for safer anesthesia delivery. We should stop being cursed by the need for “MRI-compatibility” and start actively implementing an “anesthesia-compatible” MRI environment. To achieve this goal, anesthesiologists should be involved from the beginning of the conceptual design of MRI suites.

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In Reply.—We appreciate the interest of Miyasaka et al. in our recent letter1 and applaud them for their comments. Miyasaka et al. demand the development of anesthesia-compatible magnetic resonance imaging (MRI) suites and monitoring techniques because conditions to provide anesthesia for patients in MRI suites are often far from optimal and provoke mistakes.

In particular, the authors address difficulties achieving adequate monitoring within the MRI suite, especially in critically ill patients. Clearly, the problem is that monitors, ventilators, and infusion pumps either interfere with the MRI (evoking low quality images or artifacts) or the equipment contains ferromagnetic parts and may not work correctly in the presence of a strong magnetic field (which results in danger for the patient). At the same time demand for MRI diagnostic procedures in critically ill patients is increasing steeply. Thus, with present equipment the anesthesiologist often faces a “catch-22” situation.

Accordingly, Miyasaka et al.’s points that MRI manufacturers should take anesthesiologists’ comments regarding more comfortable and safe MRI suites seriously and that anesthesiologists should be involved in conceptual designs of MRI suites from the beginning are well

References


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Cotê and at least temporarily adopted by Dr. Kawabata's institution, is particularly worthy of attention is Miyasaka. The form of asking manufacturers to be anesthesia-compatible at the industry thinking such that there is harmony and balance in procedures for anesthesia care in MRI suites and for orienting (reorienting?) is sought only after an MRI suite has been constructed and procedures have been streamlined for use by patients who are neither sedated nor awake (invasive ventilation?). The letter by Miyasaka et al. is highly relevant to such incidents because of the very important points it makes regarding safety violations during clinical anesthesia in the magnet room. Miyasaka et al. note that in many instances, input from anesthesiologists is sought only after an MRI suite has been constructed and procedures have been streamlined for use by patients who are neither sedated nor anesthetized. Strong support is appropriate for Miyasaka et al.'s call for early involvement by anesthesiologists, both in establishing hospital procedures for anesthesia care in MRI suites and for orienting (reorienting?) industry thinking such that there is harmony and balance in the form of asking manufacturers to be anesthesia-compatible at the same time that anesthesiologists are asked to be MRI-compatible. Particularly worthy of attention is Miyasaka et al.'s point that hospital managers, contractors, and equipment managers have no professional guidelines or standards regarding the design and construction of anesthesiology facilities inside the magnet room of an MRI suite. The American Society of Anesthesiologists has produced the Operating Room Design Manual and a booklet on setting up safe office-based anesthesia, but these do not address key MRI safety issues.* Advances in magnet technologies and medical imaging are likely to require that anesthesiologists function in environments having stronger magnetic field gradients (which exert mechanical force) and more intense radiofrequency power (an energy source with the potential for burns) than one currently encounters. Anesthesiologists undergo training and make considerable efforts to learn about MRI-related dangers, and they and the patients deserve anesthesia-friendly, patient-safe systems in MRI suites. But there is truth in the title of Chester L. Karras' widely known book "You Don’t Get What You Deserve, You Get What You Negotiate." In pointing out the need for MRI suite anesthesia standards Miyasaka et al. are really calling for help in negotiating what is deserved. What should be the form? One solution would be a comprehensive statement sanctioned by the appropriate American Society of Anesthesiologists committees, possibly involving other relevant professional groups. If there are manufacturers who do not find anesthesia safety issues to be "compelling," then anesthesiologists need to provide better, more persuasive arguments, as there can be no compromise on safety. I am impressed by the altruism I have seen in the technical people I have met who are associated with medical manufacturers. Many, in choosing their profession, have shown that they are as highly motivated by seeing patients helped by optimum imaging technologies as surgeons and anesthesiologists are about seeing patients helped by the best invasive procedures. Altruism, however, was not something that the late Senator Everett Dirksen relied on almost half a century ago in a saying he was fond of: "When I feel the heat, I see the light." It seems appropriate that there be follow-up to the letter by Miyasaka et al. in the form of anesthesiologists getting together to provide some heat and light for themselves, hospital colleagues, and equipment manufacturers.

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Proponents of Liberalized Fasting Guidelines

To the Editor:—We read with interest the case report by Kawabata et al. of pulmonary aspiration on induction of anesthesia in an infant fed formula 4.5 h before surgery. We have long been proponents of liberalized fasting guidelines for clear liquids and, more recently, for infant formula. For infants younger than 6 months of age, expert opinion has been equally divided as to 4 h or 6 h formula fast requirements. The liberalization of formula fasting to 4 h, as outlined by Dr. Coté and at least temporarily adopted by Dr. Kawabata’s institution, is a practice we continue to support in healthy infants younger than 6 months old. We recognize that there are very limited data to support either the safety or the hazard of such a practice.

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Reference

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events in two 34-month epochs, one before and one after the liberalization of infant formula fasting from 6 h to 4 h, and found no increased incidence (table 1). In fact, for reasons that remain unclear, there was a 10-fold reduction in overall incidence of pulmonary aspiration from the first epoch to the second (Fisher exact test $P = 0.0002$.) With regard to patients <6 months of age, however, there were no documented aspiration events in any of the 9,266 infants cared for in the past 6 yr. Our experience is consonant with that of others. It is important to note that although we allowed healthy infants formula at 4 h before procedures from July 2001 on, we have no data as to how many infants actually fed between 4 h and 6 h before induction of anesthesia. Lastly, from a more global perspective, if we were to assume that the 50% of North American and European practitioners surveyed by Emerson et al. and Ferrari et al. allow a 4-h fast and have not found an increased incidence of pulmonary aspiration, then the evidence for the safety of this practice may be growing.

As for the case presented by Kawabata et al., she appeared to be a healthy infant who came for elective surgery of the lip. Height and weight appeared normal, consistent with normal gastrointestinal function and nutritional status. Nevertheless, undiagnosed chronic and subclinical acute disorders may have caused gastrointestinal dysmotility that could have contributed to the aspiration described. Both total feed volume (260 ml within 1 h) and formula characteristics (formula derived from cow milk) may have contributed to the event as well. A 120-ml feed might be more typical and appropriate 4 h before anesthesia and surgery in a young infant. In our fasting study, feed volumes averaged 120–150 ml. Human milk and whey-based formula are emptied faster than casein-based formula and cow’s milk. There is insufficient data to state with certainty that one formula is safer than another, but given the slower emptying times for the latter, a case can be made for either avoiding them or prolonging the fast.

Kawabata et al. misquoted our GFV study by stating “...9% of formula-fed infants who fasted for 4 h had undigested traces of formula in their gastric content.” In fact, 9% (9/97) of all of our subjects had traces of residual formula. Evidence of formula was found in 13% of subjects fasted for <6 h and in 3% of those fasted for ≥6 h. Interestingly, one 8.5-month-old infant who fasted for 10 h still had a white tinge to the recovered gastric aspirate. We believe that these white-tinted residua associated with small GFV and pH not different from traditional fasts do not significantly increase pulmonary aspiration risk. Given the limited data, however, practitioners who wish to reduce the risk of even trace amounts of formula in the stomach may feel more comfortable recommending a 6-h fast. Finally, GFV remains an imperfect surrogate marker for pulmonary aspiration. Small GFVs do not preclude vomiting and aspiration of upper small bowel contents via retrograde giant contractions and no study has examined small bowel emptying time as it relates to formula feeds.

The central issue is that pulmonary aspiration of gastric contents is a rare event. Mortality or significant morbidity associated with pulmonary aspiration is exceedingly rare. To carefully measure the impact of various fasting regimens on even uncomplicated pulmonary aspiration, randomized, controlled trials would require sample sizes in excess of 30,000. In the past 6 yr, overall patient volume at The Children’s Hospital of Philadelphia exceeds this number but not in the infant subpopulation of interest. No matter how fasting guidelines are designed, the risk of pulmonary aspiration of gastrointestinal contents, even in fasted healthy infants, will never be zero. None of this diminishes the importance of the case report. When events are rare, the case report may be the only way of tracking emerging patterns and trends that will enhance our understanding and permit best practice to continue to evolve.

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To the Editor.—Kawabata et al. reported a case of unanticipated vomiting and pulmonary aspiration after induction of anesthesia in a formula-fed 4-month-old female infant who weighed 6.26 kg scheduled for elective cheiloplasty. Anesthesia was induced with sevoflurane and nitrous oxide in 33% oxygen. The anesthesiologist tried to maintain spontaneous respirations but, abruptly, mask ventilation could not be performed. Arterial oxygen saturation decreased to 28% and formula was found in the patient’s mouth. The authors argue that despite surveys and the American Society of Anesthesiologists practice guide-
appears from their own description that the action was regurgitation rather than vomiting. Various factors can play a role in inducing regurgitation in infants. The Pediatrician should proceed with the assumption that the stomach, encroachment of some abdominal organs, previous air swallowing during crying, and diaphragmatic breathing. In addition, a mild from of relaxation of the gastroesophageal junction may exist in the first 6 months of age. Once gastric contents are forced up into the esophagus, they may readily find their way into the pharynx because of the short esophagus in the infant.

Third, although inhalation induction using 30–35% oxygen is commonly practiced in infants and children, it is time to seriously consider “preoxygenation” to increase oxygen reserves and thereby delay the onset of arterial oxygen desaturation in case airway obstruction occurs. Infants are at an increased risk of hypoxemia because of their small functional residual capacity and increased oxygen consumption. The value of preoxygenation in pediatric patients has been recently discussed in a number of reports and certainly can be easily achieved.

In conclusion, despite what appears to be an adequate preoperative fasting, the anesthesiologist should proceed with the assumption that the stomach is not completely empty. It behooves the anesthesiologist to take into consideration the factors that may induce regurgitation during induction and specifically avoiding and immediately correcting airway obstruction. Finally, preoxygenation before induction should be practiced.

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In Reply—First of all, we would like to convey our appreciation to Dr. Cook-Sather and Dr. Salem for their scientific and detailed responses to our Letter to the Editor.

As Dr. Cook-Sather rightly pointed out, although studies have been conducted, and scientific data accumulated before establishing the guidelines for fasting periods, there is very limited data to definitively support a 4h versus a 6h fasting period. We are of the opinion that the presence of guidelines for two different fasting times could prove problematic. After encountering this case, we changed our fasting time policy from 4h for formula milk to 6h, as we thought that it provided a greater safety margin for the risk of aspiration while not causing any serious disadvantage to the patients and their parents.

Dr. Salem commented that despite an adequate fasting period, infants, owing to their anatomical and physiologic differences from adults, are predisposed to regurgitation of gastric contents. Hence, they should be treated as potential risks for regurgitation and all precautionary measures to minimize this risk should be undertaken. We are in complete concurrence with this opinion.

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(Accepted for publication September 1, 2004.)
To the Editor—In their article about pacemakers and extracardiac radiofrequency ablation, Tong et al. describe some appropriate precautions; i.e., have temporary equipment available for backup pacing and check the pacemaker with a programmer both before and after the radiofrequency ablation event. However, the authors should have included a statement about the placement of the current return pads, which should be as close as possible to the radiofrequency ablation delivery electrodes to prevent or minimize radiofrequency ablation current travel across the generator and lead system (they did not specify the location of these pads). In addition, this article has a number of inaccuracies.

First, the authors have misinterpreted electrical artifacts displayed by their digital monitor (and marked with upward arrows in strips D and E [fig. 1]) as pacemaker output pulses, making an incorrect diagnosis of “intermittent temporary runaway pacing.” None of these pulses appears to “capture” the ventricle and pace the heart, and other pacing pulses exist in an appropriate fashion that do depolarize the ventricle. In fact, these are pacemaker “pseudospikes,” which result from inappropriate digital processing of the electrical noise from the radiofrequency ablation by the electrocardiographic monitor. This phenomenon has been well described. Additional pseudospikes can be seen in strip D after complexes 5 and 8 and in strip E after complexes 2, 6, and 7. Also, the term “runaway pacing” refers to continuous, high-rate pacing resulting from internal component failure within a generator. Correction is always by pacemaker replacement. Further, these pseudospikes appear at rate of 375 bpm (strip E, 160 ms), arguing against runaway pacing, as the Guidant Meridian pacemaker (Guidant Corporation, St. Paul, MN) has a runaway limit of 205 bpm.

Next, the authors report that their patient had “complete atrioventricular block.” Complete atrioventricular block is not demonstrated by any of the five strips that accompany the text. Strip C and strip E (complexes 4–7) show narrow complex QRS complexes, suggesting that the patient had intact atrioventricular nodal conduction. The rhythm might be sinus at approximately 70 bpm with a PR interval of 300 ms. It is not clear if the atrial events are paced, as operating room monitors do a poor job of detecting and showing atrial pacing events. Because the authors did not describe the programmed PR interval, we have no way to know whether atrial events are paced or native. The atrioventricular delay in strip D and possible paced atrial events in strip A, complexes 8–10 (open downward arrows), do appear to be less than 200 ms. However, without knowing the atrioventricular delay programming and without real-time data from the pacemaker, limited conclusions can be made about the longer atrioventricular delays seen throughout the five strips.

The authors state that the patient’s heart rate immediately changed from 63 to 96 bpm with the onset of radiofrequency ablation delivery, seen in strip A complexes 4–7. Yet, there is the suggestion of P waves (open downward arrows, added) at these complexes. The possible atrial pacing artifacts at complexes 6–10 (strip A) might have resulted from rate smoothing, which will limit any decreasing rate in this pacemaker, if programmed. Thus, without knowledge of the pacemaker settings as well as real-time data from the pacemaker (telemetry), no conclusion can be reached about the etiology of this increased heart rate.

At most, these strips might demonstrate atrial and ventricular oversensing, which should be expected during radiofrequency ablation, according to the Meridian Physician’s Manual. In a DDD-programmed pacemaker, the occurrence of atrial oversensing, without ventricular oversensing, will result in higher rates of ventricular pacing than expected; usually, these events take place at the upper limit for tracking. With the presumed upper limit for tracking in this case of 120 bpm, atrial oversensing could have taken place at A4–6 and B5–6. Ventricular oversensing is difficult to prove with these strips, as the longest RR interval occurs at A9–10 and represents a heart rate of 70 bpm. In the setting of ventricular oversensing, the pacemaker will fail to pace the ventricle and an inappropriately long RR interval will occur. No such intervals are demonstrated.

In summary, Tong et al. remind us that extracardiac radiofrequency ablation in a patient with a cardiac generator should be approached with caution owing to possible electromagnetic interference with the generator and potential misinterpretation of electrocardiographic behavior.

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In Reply.—In our letter, we only noted that the radiofrequency current interfered with the function of the pacemaker but that there was no damage the pacemaker itself. Actually, we described this effect as “falsely inhibited,” which was very similar to what was first described by Chin et al. Chin et al. noted that intracardiac radiofrequency current may produce a number of potentially serious pacing systems malfunctions. When such ablation was performed in close proximity to a pacing lead, false inhibition was observed, even when devices were programmed to the asynchronous mode. Several devices paced at abnormal rates during current flow, producing extremely rapid pacemaker runaway. Some pacers reverted to a noise mode of operation during the ablation procedures. Despite these effects, none of the devices was spuriously reprogrammed by the ablation, with the exception of revision to reset mode. Nor were any of the pulse generators permanently damaged. In our case, the situation was similar and that is why we described the device as “falsely inhibited” rather than “true inhibition” in our first submission. However, some reviewers had different opinions and were uncomfortable with the term “falsely inhibited.” They insisted that the generator is either inhibited or it is not. Finally we omitted the word “falsely.”

Second, a statement is needed regarding the placement of the current returns pads, which should be as close as possible to the radiofrequency ablation delivery electrodes to prevent or minimize radiofrequency ablation current travel across the generator and lead systems. In this case, two grounding pads were applied to the posterior aspects of the patient’s thighs as close as possible to the ablation electrodes. This comment was deleted during the revision of the work.

Third, complete supra-Hisian Atrio-ventricular block was diagnosed by a cardiologist via a formal electrophysiological study.

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(Accepted for publication September 3, 2004.)

To the Editor:—The July 2004 issue of Anesthesiology contained a report by Estebe and Myers demonstrating neurotoxic effects of amitriptyline when injected in high doses immediately adjacent to the rat sciatic nerve. Local anesthetic properties have been demonstrated with high doses of amitriptyline, and local anesthetics also have been shown to produce neurotoxicity after similar methods of administration. The demonstration of neurotoxic effects following local administration is an important reminder of the need for careful assessment of novel routes of drug administration.

Unfortunately, there is a significant error in the calculation of the total dose administered in the Estebe and Myers study. Thus, they inject 0.2 ml of 25 mg/ml or 79.6 ml amitriptyline as their highest dose and compute this to correspond to a dose of 16 nmol. However, this dose actually corresponds to a dose of 16 μmol; therefore their dose computation is in error by a factor of 1000 (table 1, section A). The doses administered are comparable to those administered in demonstrating local anesthetic properties of amitriptyline (table 1, sections B1, B2) but are significantly higher than those administered in producing antinociception (table 1, section C). Much of their discussion regarding amitriptyline systematically perpetuates the thousand-fold error in calculations, and this results in some very misleading considerations.

After peripheral administration, by injection locally into the dorsal or plantar surface of the hindpaw, amitriptyline produces antinociception in rat models of ongoing (formalin test) or neuropathic pain (spinal nerve ligation model). Efficacy of locally administered amitriptyline has also been demonstrated in a rat model of diabetic neuropathy. In each of those studies, doses of 100 nmol amitriptyline produced antinociception against spontaneous (formalin) and evoked (thermal, mechanical) behaviors. This dose, however, had no effect on thermal latencies in the uninjured state and did not produce tissue edema. Increasing the dose of amitriptyline to 300 and 1000 nmol leads to increases in thermal thresholds in the uninjured state, which may reflect involvement of local anesthetic properties. These higher doses also cause tissue edema that, at the 1000 nmol dose, persists to some degree at 24 h, but mechanisms involved in edema are unclear.

The Estebe and Myers study evaluates doses of 2,000–16,000 nmol amitriptyline injected immediately adjacent to the sciatic nerve in rats. When amitriptyline up to 100 nmol is administered locally into the hindpaw of rats in models of antinociception, it is administered at a much lower total concentration and into tissue that is not necessarily in the immediate vicinity of the nerve. There is no data to suggest that neurotoxic or overt local anesthetic properties are involved in antinociception at doses up to 100 nmol. Indeed, the actions of amitriptyline at these doses are substantially blocked by methylxanthine adenosine receptor antagonists, and it is very unlikely that such antagonists would block neurotoxic or local anesthetic actions. This issue is raised in the context of implicating a receptor-operated mechanism in the action of amitriptyline rather than in the context of implicating any

The Estebe and Myers study demonstrates that amitriptyline is a potent local anesthetic at doses that are comparable to those used in antinociceptive studies. However, the doses used in the antinociceptive studies are much lower and are not sufficient to cause tissue edema. Therefore, the actions of amitriptyline in the antinociceptive studies are not likely to be due to local anesthetic properties.

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single particular mechanism in its action. Thus, amitriptyline is a complex drug with a range of pharmacological actions (given acutely it blocks noradrenaline, 5-HT, and adenosine uptake, inhibits α-adrenergic, histamine H1, 5HT2 and N-methyl-D-aspartate receptors, and blocks Na+, Ca2+, and even K+ channels), and many of these effects could contribute to peripheral antinociception. Clearly, any form of topical application of amitriptyline to the skin in humans will need to proceed with due caution regarding the potential for local toxicity.

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In Reply—We thank Dr. Sawynok for the interest in our study1 and for correcting the error in the conversion of our doses from mM to nmol. Fortunately, the experimental protocol, preparation and dosing of drug, and results and conclusions of our article remain unchanged, as this error was only one of unit conversion for the reader. The error does not affect the experimental paradigm or significance of the results. We also thank Dr. Sawynok for comparing the dose used in several studies of amitriptyline in their table, as this helps place our experiments in context. The range of dose evaluated in our study was in accordance with the doses previously used in studies that reported a prolongation of sciatic nerve blockade2 (i.e., 5–0.625 mg or 16,000–2,000 nmol; each dose administered in a 0.2-ml volume adjacent to rat sciatic nerve). Our data clearly demonstrate that amitriptyline causes a dose-related neuropathologic change if it is administered in the immediate vicinity of a nerve, and suggests that it should not be used as a local anesthetic agent. This conclusion remains unchanged. Moreover, it has recently been reported in a volunteer study that amitriptyline (25.2 mg to 6.3 mg) did not provide better ulnar nerve blockade at the wrist level than current local anesthetics.3 Indeed, amitriptyline has a reduced margin of safety and an increased potential for neurotoxic injury.

We did not test the neurotoxic effect of a 100 nmol dose used by Sawynok et al.,4 and Esser and Sawynok5 because that dose is not associated with neural blockade of major nerve bundles. A 100-nmol dose produces an analgesic effect of uncertain mechanism, as Dr. Sawynok notes. However, the point should be made that local neurotoxic injury is concentration-dependent and that there may be subclinical neurotoxic injury with even small doses of high concentration solutions. Although the analgesic effect of amitriptyline is of considerable interest, it should also be noted that adverse effects were reported when slightly higher doses (0.3 ml of 500 nmol) were administered transdermally6 or by subcutaneous injection (300 to 1000 nmol).7

Thus, we believe there is consensus that the use of amitriptyline as a local agent in clinical situations for analgesia or anesthesia requires additional experimental study that includes a neuropathologic and a behavioral or electrophysiologic endpoint. We support these studies and regret that our conversion error may have introduced some confusion in the literature.

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The Macintosh Laryngoscope Blade

To the Editor—Burkle et al.1 perpetuate the belief held by younger practitioners of the specialty that Robert Macintosh added the curve to his laryngoscope blade to “lessen the chance of damage to the patient’s upper teeth.” Although Sir Robert indicated that exposing the larynx with a long straight blade “occasionally jeopardizes the patient’s upper teeth or takes a minor divot out of the posterior pharyngeal wall,”2 his primary purpose was to facilitate exposure of the vocal cords. Avoidance of damage to the teeth or soft tissue was not his primary purpose. Having observed a pediatric anesthetist use a straight blade in much the same manner as we now use the Macintosh blade, he modified the blade to allow the tip to “fit into the angle made by the epiglottis and the base of the tongue.” Elevation of the laryngoscope pushes the base...
of the tongue upward whereas the epiglottis, because of its attachment to the tongue, is drawn upward, providing a clear view of the larynx.

Of greatest import, however, was Macintosh’s observation, confirmed 2 months later by Rowbotham,\(^7\) that it was possible to easily expose the larynx at a lighter plane of anesthesia than with any of the standard blades. Before the ubiquitous use of neuromuscular blocking agents to facilitate intubation and of intravenous agents and halogenated hydrocarbons to induce and maintain anesthesia, relatively deep anesthesia was required to avoid laryngospasm when the epiglottis was elevated because the glottis and the inferior surface of the epiglottis are innervated by the vagus nerve. When the use of cyclopropane became commonplace, bradycardia and hypotension, attributed to a vagal-vagal reflex when the epiglottis was lifted with a straight blade, was said to occur with alarming frequency. This untoward event, too, could be avoided with the use of Macintosh’s blade because the superior surface of the epiglottis and the vallecula are innervated by the glossopharyngeal nerve.

Over the years, there have been a number of modifications of the Macintosh blade\(^4\) that purportedly reduced the danger of damage to the upper incisors and facilitated exposure of the larynx. None have met with even a modicum of success. Although Sir Robert’s blade is still one of the most popular, contemporary anesthetic techniques have eliminated its original purpose—intubation of the trachea under light anesthesia.

**References**


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To the Editor—Correct localization of an implanted infusion pump injection port only by palpation can sometimes be cumbersome, especially in obese patients and when there are no visible signs of previous successful punctures. Fluoroscopy, if available, is sometimes used in these difficult cases. We found that ultrasound can be helpful to facilitate puncture and that the method is easily practicable.

A 43-yr-old patient with an implanted intrathecal drug delivery device (Archimedes Implantable Constant-Flow Pump; Codman, Raynham, MA) was suffering from acute breakthrough pain of malignant origin (duodenal carcinoma with lumbar bone metastases). We decided to deliver a bolus through the bolus injection port of the implanted pump. This type of pump has a separate bolus port located on the outer end of the circular symmetrical body, distant from the centrally located port to refill the reservoir chamber. Thus, depending on the orientation of the pump during the implantation, the bolus port can be found anywhere on the circumference and may be difficult to localize depending on the overlying tissue. Moreover, because aspiration through a correctly positioned bolus needle is not consistently feasible, control possibilities after positioning of the needle are limited. In our case, repeated careful palpation of the skin to find the port was inconclusive. Therefore, we decided to use sonography for localization. With a 5–10 MHz linear “hockey-stick” transducer attached to a portable ultrasound device (SonoSite 180; SonoSite, Bothell, WA) we could easily visualize the bolus port (fig. 1). The skin was marked accordingly and a needle was introduced successfully at this point perpendicular to the skin.

Only in one case report by Egerszegi et al. in 1990 was ultrasound used to facilitate the repeated localization of a soft expander injection port in a pediatric patient. It is important to note that we did not perform real-time guidance of the needle under ultrasound in this case but only used sonography to mark the puncture site before disinfection and insertion of the needle. It seems to be easier for this application, is effective, and avoids sterile wrapping of the transducer.

Ultrasound guidance has gained increasing interest in regional anesthesia and pain medicine in recent years. Many private offices and outpatient pain clinics and most hospitals are equipped with ultrasound devices today. The development of smaller, portable systems has further increased the availability of ultrasound. Compared with fluoroscopy that could also be used to facilitate port localization in difficult cases, ultrasound is portable today, more easily available, and not associated with exposure to ionizing radiation. We believe that with this easy-to-learn method, which is another useful application of ultrasound in pain medicine, multiple puncture attempts can be avoided when conventional localization of a pump injection port becomes difficult.

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