Verapamil Worsens Rate of Development and Hemodynamic Effects of Acute Hyperkalemia in Halothane-anesthetized Dogs: Effects of Calcium Therapy

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The hemodynamic effects of verapamil pretreatment versus no pretreatment were evaluated in five acutely hyperkalemic dogs. Using ECG evidence for severe hyperkalemia, the halothane-anesthetized dogs were rendered acutely hyperkalemic to similar plasma levels of K⁺ (K⁺ = 8.2 ± 0.8 mEq/l verapamil plus hyperkalemia, K⁺ = 9.4 ± 0.2 mEq/l hyperkalemic controls). The verapamil-hyperkalemic group had significantly lower cardiac indexes (CI) (CI = 1.3 ± 0.5 l·min⁻¹·m⁻² verapamil plus hyperkalemia vs. CI = 3.0 ± 0.2 l·min⁻¹·m⁻² hyperkalemic controls) and lower mean arterial pressures (MAP = 60 ± 13 mmHg verapamil plus hyperkalemia vs. MAP = 96 ± 7 mmHg hyperkalemic controls). Calcium therapy for hyperkalemia that returned CI to control levels in hyperkalemic controls only partially reversed the severe hemodynamic depression and did not improve the AV block seen during hyperkalemia in the presence of the calcium entry blocker verapamil. Surprisingly, the total mEq of KCl infused at the same rate into verapamil-pretreated dogs to result in similar high serum potassium levels was only one-third that required in dogs not pretreated with verapamil (1.6 ± 0.3 mEq/kg KCl in verapamil–hyperkalemic group vs. 5.0 ± 0.7 mEq/kg KCl in hyperkalemic controls). The authors conclude 1) verapamil renders hyperkalemia likely after much less intravenous K⁺ administration; 2) the hemodynamic depression seen during acute hyperkalemia in halothane-anesthetized dogs is much more severe in the presence of verapamil; 3) calcium therapy is only partially effective in reversing the hemodynamic depression caused by hyperkalemia in the presence of the Ca⁺⁺ entry blocker verapamil; 4) Ca⁺⁺ in the dosage studied was not protective for second-degree A-V block seen in the acutely hyperkalemic dog pretreated with verapamil and anesthetized with halothane. (Key words: Anesthetics, volatile; halothane. Heart: anti-arrhythmia agents, verapamil; myocardial function. Ions: calcium; potassium, hyperkalemia.)

VERAPAMIL INHIBITS slow inward calcium currents across excitible membranes.¹ The drug recently has been approved in the United States for intravenous treatment of supraventricular tachyarrhythmias and orally for angina pectoris. Increasing numbers of surgical patients now are receiving verapamil.

Verapamil¹⁻³ and halothane⁴⁻⁸ are both direct myocardial depressants and verapamil¹⁻³ also decreases peripheral vascular resistance. Verapamil,²,⁷,⁸ high concentra}

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Methods

Two separate experiments were done at least one week apart on each of five fasting mongrel dogs weighing 10.7 ± 0.4 kg (mean ± SE) and anesthetized with steady state 1 MAC end-tidal halothane (Beckman® LB II analyzer; Beckman Instruments Inc., Fullerton, California). Succinylcholine (30 mg) was used to facilitate tracheal intubation and thereafter was infused continuously (100 mg/h) to provide muscle paralysis. Mechanical ventilation was maintained and PaO₂ was held as close to 150 mmHg as possible by adjusting inspired O₂ and N₂ concentrations. Sodium bicarbonate was administered if buffer base was below 40 mEq/l.

Separate peripheral intravenous cannulas were placed for drug and fluid administration. The left common carotid artery was cannulated for blood sampling and for monitoring of mean arterial pressure (MAP). Using a left external jugular vein, a 5 Fr. catheter was placed in the pulmonary artery for measuring right atrial (RA), pulmonary artery (PA), pulmonary artery wedge pressure (PAWP), and thermal dilution cardiac output (CO), the latter with an Instrumentation Laboratories 701 Cardiac Output Computer® (Instrumentation Laboratory Inc., Lexington, Massachusetts). EKG was recorded on a Grass 810® recorder (Grass Instrument Co., Quincy, Massachusetts).
### Table 1. K⁺, Ca²⁺, and Hemodynamic Measurements in Dogs

<table>
<thead>
<tr>
<th></th>
<th>Serum K⁺ mEq/l</th>
<th>Whole Blood Ionized Ca²⁺ mEq/l</th>
<th>HR (BPM)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Group 1</td>
<td>Group 2</td>
<td>Group 1</td>
</tr>
<tr>
<td>Control</td>
<td>3.3 ± 0.1</td>
<td>3.5 ± 0.3</td>
<td>2.7 ± 0.1</td>
</tr>
<tr>
<td>Postverapamil</td>
<td>9.4 ± 0.2*</td>
<td>8.2 ± 0.8*</td>
<td>2.9 ± 0.2</td>
</tr>
<tr>
<td>Prior to Ca²⁺ therapy</td>
<td>3.3 ± 0.2*</td>
<td>9.3 ± 2*</td>
<td>3.6 ± 0.2*</td>
</tr>
<tr>
<td>5 min post Ca²⁺ therapy</td>
<td>7.4 ± 0.5*</td>
<td>6.6 ± 0.7*</td>
<td>3.1 ± 0.1</td>
</tr>
</tbody>
</table>

N = 5, mean ± SEM

* Significant difference from control.
† Significant difference between Group 1 and Group 2.
‡ Between 1 and 5 min after calcium administration, three of five

Plasma (K⁺) and whole blood ionized Ca²⁺ were measured with a calibrated Clin-Ion® (Applied Medical Technology, Inc., Palo Alto, California) Cl⁻ ion sensitive electrode. Urine production and urine (K⁺) also were measured in three animals. Plasma verapamil and norverapamil were measured by a modification of the procedure of Harapat and Kates. Verapamil, norverapamil, and an internal standard (D-517 from Knoll Pharmaceutical Company, Wippany, New Jersey) were extracted from alkalinized serum (4:1 v/v, plasma 0.5 M NaOH) into diethyl ether. The drugs were then back-extracted into a small volume of dilute nitric acid (pH 2.5). The extract was chromatographed on a Beckman 5-μm, 15 cm × 4.6 mm ODS column with a mobile phase consisting of 30% acetonitrile in 0.1 M lithium phosphate containing 1% triethylamine. The endogenous fluorescence of each molecule was measured (ex. λ = 203 nm, em. λ = 320 nm) and used to quantitate each. Day-to-day reproducibility (N = 16) of this procedure was 4.7% for verapamil and 3.6% for norverapamil at 100 ng/ml.

After control hemodynamic and blood gas measurements were obtained, animals were given iv KCl, infused at 0.075 mEq·kg⁻¹·min⁻¹. During one of the two separate studies in each dog, no verapamil was given (Group 1). In the other study, in each animal, verapamil 0.15 mg/kg bolus, plus a verapamil infusion of 5.6 μg·kg⁻¹·min⁻¹ was given 10 min prior to beginning the KCl infusion (Group 2). The two studies in each dog were separated by at least 1 week and were performed in random order. Hemodynamic and blood gas measurements were repeated 10 min after the bolus of verapamil was given and every 10 min after the KCl infusion was begun. KCl was infused until ECG changes typical of severe hyperkalemia were seen. These consisted of tall peaked T waves, widened QRS, loss of P waves, "sine wave" QRS configuration, or severe second degree AV block. Prior to calcium administration, hemodynamic recordings and blood samples for K⁺ and verapamil determinations were collected. Calcium chloride (14 mg/kg) was given as a rapid bolus and the KCl infusion was stopped. Hemodynamic measurements, blood–gas, and K⁺ determinations were obtained 1, 3, 5, 15, and 30 min after calcium administration.

Statistics were by Student’s t test for paired and unpaired values as appropriate with P < 0.05 considered significant.

### Results

During the study, PaCO₂ averaged 39 ± 1 mmHg (mean ± SE), PaO₂ 151 ± 4 mmHg, buffer base 39 ± 0 mEq/l, and end-tidal halothane 0.79 ± 0.01%

Verapamil, studied 10 min after intravenous administration but prior to K⁺ infusion, did not result in significant changes in cardiac indexes (CI), MAP, PAPW, SVR, or HR (table 1). Group 1 (hyperkalemia alone) and Group 2 (verapamil plus hyperkalemia) were rendered acutely hyperkalemic to similar levels of serum K⁺ prior to Ca²⁺ therapy. (Group 1 K⁺ = 9.4 ± 0.2 mEq/l and Group 2 K⁺ = 8.2 ± 0.8 mEq/l).

Hyperkalemia in both Group 1 and Group 2 resulted in decreased CI compared with controls (Group 1 control CI = 4.2 ± 0.5 l·min⁻¹·m⁻² decreased to 3.0 ± 0.2 l·min⁻¹·m⁻² and Group 2 control CI = 3.7 ± 0.5 l·min⁻¹·m⁻² decreased to 1.3 ± 0.5 l·min⁻¹·m⁻²). The decrease in CI from control values during hyperkalemia was significantly greater in verapamil-pre-treated dogs (Group 2 65% decrease in CI vs. Group 1 29% decrease in CI).

One minute after Ca²⁺ therapy, CI returned to control in Group 1 (control CI = 4.2 ± 0.5 l·min⁻¹·m⁻² vs. 4.2 ± 0.4 l·min⁻¹·m⁻² 1 min after Ca²⁺). In Group 2, CI was still significantly depressed 1 min after Ca²⁺ compared with control (Group 2 control CI = 3.7 ± 0.5 l·min⁻¹·m⁻² vs. CI = 2.4 ± 0.4 l·min⁻¹·m⁻² 1 min after calcium therapy). The difference in CI between Group 1 (4.2 ± 0.4 l·min⁻¹·m⁻²) and Group 2 (2.4 ± 0.4 l·min⁻¹·m⁻²) 1 min after Ca²⁺ is significant.

A decrease in MAP was seen only in the verapamil-
pretreated hyperkalemic animals (Group 2 control MAP = 94 ± 13 mmHg vs. 60 ± 13 mmHg prior to Ca++ therapy and 66 ± 7 mmHg 1 min after Ca++ therapy).

Heart rate was lower in the verapamil-pretreated animals with hyperkalemia 5 min after Ca++ therapy (Group 2 HR = 86 ± 12 beats/min vs. Group 1 HR = 154 ± 16 beats/min).

Second degree A-V block was seen in three of five animals in both Groups 1 and 2 during hyperkalemia prior to Ca++ therapy. Despite this, at 1 min after Ca++ therapy, none of the Group 1 (without verapamil) animals continued to show second degree A-V block, whereas with verapamil pretreatment, all five animals with hyperkalemia showed persistent second degree block. One minute after Ca++ therapy, the whole blood ionized calcium rose similarly in the hyperkalemic controls (2.9 ± 0.2 to 3.6 ± 0.2 mEq/l) and the verapamil pretreated animals (2.4 ± 0.1 to 3.3 ± 0.3 mEq/l). The Ca++ rise in the verapamil pretreated group, though, did not reach statistical significance.

Despite identical infusion rates, the total mEq of KCl infused into the verapamil-pretreated dogs, which was required to produce similar ECG changes and which (in retrospect) resulted in similar serum potassium levels, was only one-third that required in dogs that were not pretreated with verapamil (Group 2 1.6 ± 0.3 mEq/kg KCl administered vs. Group 1 5.0 ± 0.7 mEq/kg KCl administered). The total K+ infusion time for verapamil pretreated dogs was 21 ± 5 minutes, while 66 ± 11 minutes was required to obtain a similar level of severe hyperkalemia in controls.

In three animals, urine output and urine K+ were measured. The urine output was much greater in animals not pretreated with verapamil (Group 1 253 ± 68 ml urine with 78 ± 18 mEq K+/ml versus Group 2 (7 ± 6 ml urine with 118 ± 98 mEq K+/ml – K+ was not measured on one Group 2 animal).

One animal in Group 2 proceeded from second degree A-V block to ventricular fibrillation during severe hyperkalemia but was defibrillated successfully by DC cardioversion. In addition to the five animals reported in this article, two other animals (one without and one with pretreatment) developed ventricular fibrillation intractable to DC cardioversion during the infusion of KCl before hemodynamics could be recorded prior to Ca++ therapy.

Plasma concentrations of verapamil measured in Group 2 animals just prior to Ca++ therapy averaged 130 ± 25 ng/ml (range: 64–190 ng/ml). No accumulation of norverapamil, the principle active metabolite of verapamil, was detected in these plasma specimens. The sensitivity limit of the procedure was 5 ng/ml.

**Discussion**

The findings that HR and SVRI in halothane-anesthetized dogs were not changed significantly 10 min after intravenous verapamil administration but prior to K+ infusion (table 1) are consistent with those of Kapur and Flacke. In dogs anesthetized with 1.1 MAC halothane and given a 0.2 mg/kg verapamil bolus, they reported no significant changes in HR, and early decreases in SVRI were back to control 10 min after verapamil administration. They reported a 12% decrease in MAP that is similar to the 10% (nonsignificant) decrease seen in our study. Flacke and Kapur reported an initial increase in CI 1 min after verapamil, a decrease in CI 10 min after verapamil, and a return to control CI for the remaining 60 min of their study. Our CI was not different from control 10 min after verapamil administration. They did not study hyperkalemic animals.

Mangiardi et al. reported that at plasma verapamil concentrations of 92 ± 10 ng/ml, the drug had the desired antiarrhythmic effect, namely prolonging the atrio-ventricular bundle conduction interval and abolishing ventricular arrhythmia but that this concentration did not significantly decrease HR or cardiac output in thiopental-anesthetized dogs. Similar to the control findings of our study and the
Calcium is considered to be initial therapy of choice for acute hyperkalemia. When the resting membrane is depolarized partially by the presence of hyperkalemia, it is thought that increasing the extracellular calcium concentration brings the threshold potential also closer to zero, thus restoring previous membrane excitability albeit at less overall negative potential. In our model, Ca++ restored CI to control and eliminated second degree A-V block in hyperkalemic halothane-anesthetized animals without verapamil (Group 1). By contrast, Ca++ treatment in the presence of the above, plus the slow calcium channel blocker verapamil, only partially corrected the more severely depressed CI and had no beneficial effect on the second degree A-V block. In fact, all dogs in the verapamil-pretreated Group 2 had persistent second degree A-V block 1 min after Ca++ therapy, whereas none of the Group 1 animals had second degree A-V block after Ca++. The findings that Ca++ therapy for hyperkalemia in the presence of calcium entry blockers partially reversed the hemodynamic depression but not the A-V block is consistent with the finding by Hariman et al.,24 that increasing serum Ca++ by 40% comparable to the increase in whole blood ionized Ca++ seen in our experiment would reverse the hemodynamic depression of verapamil but not the effects of verapamil on A-V conduction, at least in normokalemic animals. High concentrations of halothane itself also are reported to decrease conduction through the A-V node. Verapamil did not appear to alter whole blood free ionized Ca++ during hyperkalemia or the rise in whole blood Ca++. Calcium therapy.

Potassium was infused into Group 1 and Group 2 dogs at the same rate to an ECG endpoint indicative of severe hyperkalemia. Near-identical peak serum K+ levels resulted. Despite this, we were surprised to find that the time and therefore the total amount of KCl infused into Group 2 dogs was explained only partially by the higher urine volume in the dogs not pretreated with verapamil (Group 1). The net potassium infused in the three Group 1 dogs which were collected was 44 mEq (63 mEq less 19 mEq excreted in the urine) compared with 16 mEq (17 mEq less 1 mEq excreted) in the three Group 2 dogs. Therefore, despite correcting for urinary excretion, almost three times as much net K+ was infused into Group 1 animals versus Group 2 in order to obtain similar ECG endpoints and serum K+ levels (determined minutes later). Decreased urine production in the verapamil–hyperkalemic group probably was prerenal in origin because of decreased MAP and CI. A possible explanation for verapamil’s apparent depressing of K+ tolerance is that verapamil may decrease net inward flux of excess K+ into cells. Plasma, interstitial, and lymph K+ are only 1.5–2% of total body K+ (≈48 mEq compared with ≈3600 mEq intracellularly).25 A major protective mechanism against hyperkalemia is movement of K+ into the large intracellular space.
cellular pool. At least in cardiac Purkinje fibers, the Ca$^{++}$ entry blockers have been shown to increase the net outward time-independent plateau current, which is largely carried by potassium ions. During hyperkalemia, calcium channel blockers similarly may decrease the net movement of excess extracellular K$^+$ intracellularly. Clearly, in our experiments, verapamil rendered the homeostatic mechanisms against development of acute serum hyperkalemia considerably (circa three times) less effective.

We conclude the following: 1) verapamil renders hyperkalemia likely after considerably less K$^+$ administration; 2) the hemodynamic depression seen during hyperkalemia in halothane-anesthetized dogs is much more severe in the presence of verapamil (plasma levels 150 ± 25 ng/ml); 3) calcium therapy (a 36% increase in whole blood ionized Ca$^{++}$) is only partially effective in reversing the hemodynamic depression caused by hyperkalemia in the presence of the Ca$^{++}$ entry blocker verapamil; 4) Ca$^{++}$ in the dosage studied is not therapeutic for second degree A-V block seen in the hyperkalemic dog pretreated with verapamil, with halothane as anesthesia.

Considering the limitation that our results were obtained in a canine model and not a human, the potential clinical significances of our findings are as follows. First, severe hyperkalemia in the patient treated with verapamil may occur after much smaller quantities of intravenous KCl and/or fewer transfusions of near-outdated blood. Verapamil also might be expected to markedly inhibit potassium tolerance in patients with chronic renal failure and inability to excrete dietary K$^+$. Second, in the verapamil-pretreated patient, more severe hemodynamic depression due to hyperkalemia will likely occur than would be expected from hyperkalemia alone. Finally, in the presence of the calcium entry blocker verapamil, although calcium therapy may improve (but not totally reverse) depressed hemodynamics due to acute hyperkalemia, it may not improve cardiac conduction defects, at least in the presence of halothane anesthesia.

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