The Antidotal Action of Thiosulfate Following Acute Nitroprusside Infusion in Dogs


The authors previously demonstrated in dogs that a bolus dose of sodium thiosulfate maintained enhanced cyanide metabolism throughout a 1-h infusion of sodium nitroprusside (SNP). To further test this antidotal action, a bolus dose of thiosulfate (150 mg·kg⁻¹) was given to eight dogs at the end of a 60-min neurethral infusion of nitroprusside (3 mg·kg⁻¹). Within 2 min of the antidote, mean plasma thiocyanate levels (70.3 μmol·L⁻¹) were significantly higher than those of seven control dogs given nitroprusside only (45.9 μmol·L⁻¹, P = 0.002) and plateaued at 153.8 μmol·L⁻¹ within 60 min, while the control values only reached 79.1 μmol·L⁻¹ (P < 0.001). Although differences between plasma cyanide levels in the two groups only attained significance 1 h after administering the antidote (0.8 vs. 2.74 μmol·L⁻¹, P = 0.03), red blood cell cyanide concentrations were significantly lower in the antidote group within 5 min (186 vs. 225 μmol·L⁻¹, P = 0.004) and remained so throughout the 2-h observation period. Compared with the controls, there was an impressive reduction in mean half-lives of plasma cyanide (25.1 vs. 74.1 min) and red blood cell cyanide (22.4 vs. 203.6 min). Similarly, peak cyanide levels occurred much sooner following the antidote (mean times: plasma cyanide 2.9 vs. 5.9 min; red blood cell cyanide 0.25 vs. 11 min). Whereas mean blood lactate levels remained elevated for at least 1 h after infusion in the controls, they showed a decrease within 5 min in the antidote dogs, becoming significantly lower 25 min later (P < 0.01). These results suggest that thiosulfate alone is a sufficient antidote for cyanide toxicity arising from excessive SNP administration. The data also suggest that red blood cells may be involved in the detoxification process. (Key words: Anesthetic techniques: hypotension, induced; nitroprusside. Enzymes: 3-mercaptopyruvate sulfurtransferase; rhodanese. Pharmacologic: nitroprusside; thiosulfate. Toxicity: cyanide; nitroprusside.)

The possibility of cyanide (HCN) intoxication during sodium nitroprusside (SNP) infusions is now well appreciated, and, accordingly, maximum doses and dose rates for acute¹⁻³ and chronic administration⁴,⁵ have been recommended. In addition, a number of studies have investigated the effectiveness of HCN antagonists in preventing toxicity when high-dose rates of SNP are unavoidable.¹⁴,¹⁶⁻¹⁹

It is generally agreed that sodium thiosulfate is the least toxic and most effective of the antidotes for prevention and treatment of HCN toxicity from SNP overdose. Studies in dogs have shown that thiosulfate, given either as a prophylactic bolus dose prior to SNP infusion⁸ or infused while SNP is administered,⁴,⁹ limits the increase in blood HCN levels. Similarly, studies in patients have demonstrated that its continuous infusion, either in parallel¹⁷ or mixed with the SNP,¹⁰ enhances HCN metabolism. Recommendations therefore have been made that whenever SNP is infused, thiosulfate always should be administered.¹⁰ However, low doses of SNP now are employed generally for hypotensive surgery, and toxicity has become a rare event. Consequently, anesthetists may well be loath to infuse another agent routinely when, in the great majority of cases, it will prove unnecessary. In addition, during chronic SNP administration, a simultaneous infusion of thiosulfate may present problems because of enhanced plasma thiocyanate (SCN) accumulation and the danger of hypovolemia.⁴ It therefore would seem preferable to give a bolus dose of sodium thiosulfate only if the SNP dose/dose rate becomes excessive.

The drawbacks of this method of administration have been claimed to be the slow onset of action and the rapid metabolism and excretion of thiosulfate.¹¹⁻¹⁵ However, our previous work⁸ suggested that these problems may have been exaggerated. It was decided accordingly to study more closely the speed and efficacy of bolus doses of thiosulfate. We therefore have given it as a postinfusion injection to dogs acutely infused with SNP (3 mg·kg⁻¹) for 1 h and have monitored changes in plasma and red blood cell HCN, and plasma SCN, using blood lactate and base deficit as indices of cellular respiration.

Methods

Anesthesia was induced with intravenous pentobarbital (60 mg·kg⁻¹) in each of 15 mongrel dogs. Endotracheal intubation was performed, and the animals then were ventilated with 60% nitrous oxide in oxygen so as to maintain arterial P<sub>CO₂</sub> between 4.6 and 5.6 kPa.
(35–42 mmHg). Incremental doses of pentobarbitone were given as required. Cannulation of the femoral artery and vein were carried out to permit blood pressure monitoring, sampling of arterial blood, and infusion of drugs. A Swan Ganz® catheter was “floated” into the pulmonary artery via a cut-down on to the external jugular vein and used for monitoring pulmonary artery pressure and for the injection of sodium thiosulfate. Each preparation was allowed to stabilize before the first (baseline) samplings and measurements were made. Arterial blood was analyzed for lactate concentrations by rapid decantation into perchloric acid and subsequent enzymatic analysis (Boehringer). Samples of plasma and red blood cells were separated from arterial blood at 4°C for immediate isolation of free HCN by the method of Boxer and Rickards,2,14 while a portion of the plasma was stored at −20°C for subsequent SCN analysis. Both HCN and SCN were measured by a modified autoanalytical technique.15 Arterial blood gas and acid–base status were determined using an IL 413 Blood Gas Analyser.®

The animals were treated randomly either as control or antidote dogs. Mean weight (±SD) of the control animals was 21.5 ± 4.4 kg and that of the antidote group was 21.3 ± 1.8 kg. Both groups received SNP (3 mg·kg⁻¹), by intravenous infusion over exactly 60 min, from a Braun infusion pump. The SNP solution was freshly prepared from the Analar® solid (Hopkin and Williams), and the total required dose was infused in a volume of 60 ml physiologic saline from a foil-wrapped syringe. The antidote group received a bolus injection of Analar® (BDH) sodium thiosulfate (150 mg Na₈S₂O₃·5 H₂O·kg⁻¹), dissolved in 5 ml deionized water, immediately upon termination of the SNP infusion.

All blood samples and recordings were taken at 20 min and 10 min before the SNP infusion, at 30 and 60 min during infusion, and at 10, 30, 60, and 120 min after the end of the SNP administration. Additional samples of blood also were taken at 2 and 5 min following SNP infusion, for lactate, blood gases, HCN, and SCN measurements. Normal saline was infused via the pulmonary artery catheter throughout the study to replace blood loss, and the animals were killed at the termination of each experiment.

**Statistical Methods**

The data from the two groups of dogs were expressed as mean ± standard error of the mean (±SEM) and compared using Student’s t test for unpaired data in the Minitab statistical package from Pennsylvania State University. A probability of 5% or less was considered significant. Half-lives (t₁/₂) of plasma and red blood cell cyanide were calculated from their log values versus time.

**Fig. 1.** Mean red blood cell and plasma cyanide concentrations in two groups of dogs during and for 2 h following a 1-h infusion of sodium nitroprusside (SNP). One group, the controls (n = 7) (——●——), received no cyanide antidote, whereas the second group (n = 8) (——O——) was given a bolus injection of sodium thiosulfate (ST) at the end of the SNP infusion.

**Results**

The changes in mean blood HCN and SCN concentrations are displayed in figs. 1 and 2, and the numeric values, SEM, and statistical significance of the differences between the two groups of dogs are given in table 1. The plasma HCN levels showed no significant differences except at 60 and 120 min postantidote (P = 0.03 and P = 0.02). However, mean red blood cell HCN levels were significantly different 5 min after the antidote was given and remained so until the end of the experiment (P < 0.01 at 65 and 180 min, P < 0.001 at 70–120...
The differences between the two groups following SNP infusion are illustrated further by the numbers of dogs in which a decrease in the measured variables had occurred at each sampling time (Tables 1 and 2) and by comparison of the times of peak HCN levels and their half-lives (t$_{1/2}$) (Fig. 3).

Correlations between the various measurements are given in Table 3. Baseline deficit showed a good correlation with blood lactate, and both were better correlated with red blood cell HCN than with plasma HCN. However, the correlations between base deficit and HCN levels in the antidote dogs were lower than in the controls, probably because of the rapid changes in HCN induced by thiosulfate. The decreases in RBC HCN levels showed a better correlation with increases in plasma SCN concentrations than did the decreases in plasma HCN. Additionally, both log plasma and log RBC HCN levels, following SNP infusion, showed a statistically significant negative linear correlation with time in both groups of dogs.

No side effects, other than transient hypotension (for 30–60 s; mean decrease in systolic pressure 8.5% ± 4.8% SEM) were seen on injection of the thiosulfate.

Discussion

Our previous study$^{8}$ demonstrated that a bolus dose of thiosulfate (75 mg·kg$^{-1}$), given 5 min prior to the start of an infusion of 1.5 mg·SNP·kg$^{-1}$ (a constant rate of 25 µg·kg$^{-1}$·min$^{-1}$ for 1 h), maintained plasma and red blood cell HCN at very low levels throughout the period of infusion, while plasma SCN concentrations climbed rapidly to reach a maximum at its termination. This contrasted with the controls where high HCN levels were reached during the infusion and plasma SCN continued to increase for 2 h after the SNP administration. These observations demonstrated that the antidotal action of a bolus dose of thiosulfate was maintained for at least 1 h and was rapid enough to facilitate the removal of HCN as it was released from the infused SNP.

In this second study, in order to further examine the speed of action of thiosulfate, dogs were infused with twice the previous dose of SNP (3 mg·kg$^{-1}$; 50 µg·kg$^{-1}$·min$^{-1}$ for 1 h) and double the dose of antidote was injected at the end of the infusion (150 mg Na$_2$S$_2$O$_3$·5 H$_2$O·kg$^{-1}$ = 12× the stoichiometric amount theoretically required to “neutralize” the HCN from the SNP dose). The SNP infusion rate employed must be close to the lethal one for dogs as indicated by the rapid increase in blood lactate and base deficit, the death of one of the control animals some 90 min after the infusion, and the fact that an equivalent dose rate of sodium cyanide has been demonstrated to be lethal within 30 min.$^{15}$
### TABLE 1. Comparison of Mean Plasma and Red Blood Cell Cyanide and Plasma Thiocyanate in Dogs Receiving No Cyanide Antidote with Those in Dogs Given Thiocyanate at the End of SNP Infusion

<table>
<thead>
<tr>
<th>Sample Time</th>
<th>Control (n = 7)</th>
<th>Antidote (n = 8)</th>
<th>P</th>
<th>Control (n = 7)</th>
<th>Antidote (n = 8)</th>
<th>P</th>
<th>Control (n = 7)</th>
<th>Antidote (n = 8)</th>
<th>P</th>
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</thead>
<tbody>
<tr>
<td></td>
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<td>μmol/L</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>-10</td>
<td>0.19 0.06</td>
<td>0.10 0.04</td>
<td>0.24</td>
<td>0.3 0.5</td>
<td>0.2 0.03</td>
<td>0.25</td>
<td>19.6 1.97</td>
<td>20.6† 2.95</td>
<td>0.78</td>
</tr>
<tr>
<td>30‡</td>
<td>3.75 0.65</td>
<td>3.42 0.21</td>
<td>0.62</td>
<td>141.1 12.59</td>
<td>115.9 12.15</td>
<td>0.58</td>
<td>51.3† 5.47</td>
<td>54.3 3.47</td>
<td>0.57</td>
</tr>
<tr>
<td>60‡§</td>
<td>5.58 0.77</td>
<td>4.65 0.51</td>
<td>0.26</td>
<td>208.1 12.93</td>
<td>209.3 10.04</td>
<td>0.92</td>
<td>45.6 4.31</td>
<td>45.9 4.03</td>
<td>0.81</td>
</tr>
<tr>
<td>62</td>
<td>5.80 0.85</td>
<td>5.00 0.59</td>
<td>0.44</td>
<td>219.8 13.30</td>
<td>202.7† 20.47</td>
<td>0.74</td>
<td>45.9 5.10</td>
<td>70.3 3.61</td>
<td>0.002</td>
</tr>
<tr>
<td>65</td>
<td>5.28 0.88</td>
<td>4.80 0.77</td>
<td>0.58</td>
<td>225.1 16.48</td>
<td>166.0 7.18</td>
<td>0.004</td>
<td>66.7 4.05</td>
<td>89.0 4.31</td>
<td>0.001</td>
</tr>
<tr>
<td>70</td>
<td>5.11 0.74</td>
<td>3.96 1.94</td>
<td>0.40</td>
<td>223.5 16.82</td>
<td>130.1 5.48</td>
<td>&lt;0.001</td>
<td>48.1 5.22</td>
<td>111.1 5.20</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>90</td>
<td>3.84 0.94</td>
<td>2.32† 0.78</td>
<td>0.25</td>
<td>229.1 25.85</td>
<td>53.1 2.16</td>
<td>&lt;0.001</td>
<td>65.1 9.05</td>
<td>145.5† 7.96</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>120</td>
<td>2.74 0.81</td>
<td>0.80 0.20</td>
<td>0.05</td>
<td>198.4 28.12</td>
<td>16.5 2.47</td>
<td>&lt;0.001</td>
<td>79.1 10.51</td>
<td>153.8 7.42</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>180</td>
<td>2.08 0.69</td>
<td>0.28† 0.09</td>
<td>0.02</td>
<td>123.6 38.55</td>
<td>7.1 0.23</td>
<td>0.006</td>
<td>112.4 12.21</td>
<td>152.4 7.07</td>
<td>0.012</td>
</tr>
</tbody>
</table>

* D = no of dogs showing a decrease in HCN level compared with the preceding value.
‡ Bolus sodium thiocyanate (150 mg·kg⁻¹) given at this point.
§ Bolus sodium thiocyanate (150 mg·kg⁻¹) given at this point.
§ One value missing.
† Sampling times during an infusion of 3 mg SNP kg⁻¹ for 1 h.

### TABLE 2. Comparison of Mean Blood Lactate and Base Deficit in Dogs Receiving No Antidote with Those in Dogs Given an Injection of Sodium Thiocyanate at the End of an Infusion of SNP

<table>
<thead>
<tr>
<th>Sample Time</th>
<th>Control (n = 7)</th>
<th>Antidote (n = 8)</th>
<th>P</th>
<th>Control (n = 7)</th>
<th>Antidote (n = 8)</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>μmol/L</td>
<td>μmol/L</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>-20</td>
<td>1.22 0.40</td>
<td>—</td>
<td>0.86</td>
<td>0.10 —</td>
<td>0.37 —</td>
<td>3.7 —</td>
</tr>
<tr>
<td>-10</td>
<td>1.16 0.37</td>
<td>—</td>
<td>0.92</td>
<td>0.09 —</td>
<td>0.52 —</td>
<td>3.7 —</td>
</tr>
<tr>
<td>30‡</td>
<td>2.55 0.38</td>
<td>—</td>
<td>2.55</td>
<td>0.17 —</td>
<td>1.00 —</td>
<td>5.94 —</td>
</tr>
<tr>
<td>60‡§</td>
<td>3.73 —</td>
<td>—</td>
<td>3.76</td>
<td>0.22 —</td>
<td>0.95 —</td>
<td>8.69 —</td>
</tr>
<tr>
<td>62</td>
<td>3.99 0.43</td>
<td>1</td>
<td>3.77</td>
<td>0.23 4</td>
<td>0.64 —</td>
<td>— —</td>
</tr>
<tr>
<td>65</td>
<td>4.09 0.43</td>
<td>4</td>
<td>5.57</td>
<td>0.21 8</td>
<td>0.28 —</td>
<td>— —</td>
</tr>
<tr>
<td>70</td>
<td>4.34 0.48</td>
<td>1</td>
<td>5.43</td>
<td>0.23 7</td>
<td>0.09 —</td>
<td>9.60 —</td>
</tr>
<tr>
<td>90</td>
<td>4.32 0.56</td>
<td>4</td>
<td>2.63</td>
<td>0.17 8</td>
<td>&lt;0.01 —</td>
<td>9.60 —</td>
</tr>
<tr>
<td>120</td>
<td>4.53 0.50</td>
<td>6</td>
<td>1.80</td>
<td>0.15 7</td>
<td>&lt;0.001 —</td>
<td>9.63 —</td>
</tr>
<tr>
<td>180</td>
<td>5.29§ 0.61</td>
<td>5</td>
<td>1.14</td>
<td>0.13 8</td>
<td>0.003 —</td>
<td>7.8§ 1.36</td>
</tr>
</tbody>
</table>

* D = no of dogs showing a decrease in level compared with the preceding measurement.
‡ Bolus sodium thiocyanate (150 mg·kg⁻¹).
§ One value missing.
† Sampling times during infusion of 3 mg SNP kg⁻¹ for 1 h.
of HCN metabolism and that it also limits its peripheral distribution in dogs.\textsuperscript{17}

Breakdown and release of HCN will continue after SNP administration ceases so that HCN toxicity may be delayed.\textsuperscript{18} This is illustrated by the control dogs where the variations in time at which peak HCN levels were reached indicate considerable individual differences in the rate of SNP breakdown (fig. 3). Also, the variation in HCN $t_{1/2}$ in the control animals shows that there are wide differences in the ability to detoxicate HCN at these high levels. No doubt this reflects variability in endogenous supplies of thiosulfate and, possibly, in the degree of inhibition of the detoxication mechanisms by elevated blood HCN.\textsuperscript{19} However, the bolus thiosulfate injection, given at the end of SNP infusion, dramatically decreased HCN $t_{1/2}$, virtually removing individual differences in detoxication rates and preventing late increases in HCN levels.

It generally is considered that HCN is converted to SCN by the action of the enzyme rhodanese (thiosulfate-cyanide sulfurtransferase, EC.2.8.1.1), which is widely distributed in the tissues and present in high concentration in the liver and kidneys.\textsuperscript{19} However, rhodanese is relatively inaccessible, being confined to the matrix of the mitochondria\textsuperscript{20} where thiosulfate only gains access slowly.\textsuperscript{19,21,22} In addition, SCN traverses cell membranes by simple diffusion\textsuperscript{23} and, therefore, after formation from HCN, would be expected to enter the plasma at a slow rate. These characteristics of the rhodanese system seem incompatible with the rapid elevation of plasma SCN in the antidote dogs. Several observations, both from this work and that reported by others, suggest the red blood cell as a possible site for HCN detoxication.

This experiment has demonstrated differences in the degree and speed of the reduction of red blood cell and plasma HCN as a consequence of an injection of sodium

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**Table 3. Correlations between Various Measurements Made in Dogs Infused with SNP (3 mg·kg\textsuperscript{-1}) for 1 h (One Group Acted as the Controls, while a Second Group Received a Bolus Injection of Thiosulfate (150 mg·kg\textsuperscript{-1}) at the End of SNP Administration)**

<table>
<thead>
<tr>
<th>Measurements Correlated</th>
<th>Control Group (n = 7)</th>
<th>Antidote Group (n = 8)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>$r$</td>
<td>$df$</td>
</tr>
<tr>
<td>RBC HCN vs. plasma HCN (0–180 min)</td>
<td>0.69</td>
<td>58</td>
</tr>
<tr>
<td>Base deficit vs. plasma HCN (0–180 min)</td>
<td>0.69</td>
<td>56</td>
</tr>
<tr>
<td>Lactate vs. plasma HCN (0–180 min)</td>
<td>0.71</td>
<td>56</td>
</tr>
<tr>
<td>Lactate vs. RBC HCN (0–180 min)</td>
<td>0.51</td>
<td>60</td>
</tr>
<tr>
<td>Lactate vs. base deficit (0–180 min)</td>
<td>0.53</td>
<td>60</td>
</tr>
<tr>
<td>Log plasma HCN vs. time (62–180 min)</td>
<td>0.55</td>
<td>47</td>
</tr>
<tr>
<td>Log RBC HCN vs. time (62–180 min)</td>
<td>0.55</td>
<td>47</td>
</tr>
<tr>
<td>Decrease in plasma HCN vs. increase in plasma SCN (62–180 min)</td>
<td>0.66</td>
<td>40</td>
</tr>
<tr>
<td>Decrease in RBC HCN vs. increase in plasma SCN (62–180 min)</td>
<td>0.86</td>
<td>40</td>
</tr>
</tbody>
</table>
thiosulfate. Thus, RBC HCN levels in the animals given the antidote were significantly lower than in the control group \((P = 0.004)\) 5 min after the antidote, whereas plasma HCN levels differed significantly \((P = 0.03)\) only after the lapse of 1 h. In the antidote group itself, the mean RBC HCN \(t_{1/2}\) was lower than plasma HCN \(t_{1/2}\) (22.4 min vs. 25.1 min). Not surprisingly, this difference was small and did not achieve statistical significance (paired \(t = 2.2, \ P = 0.064\)), since HCN passes rapidly from the plasma into the red blood cell.\(^{24,25}\) A further and important observation, previously made in humans,\(^2\) was the fact that the increases in plasma SCN, following SNP infusion in both groups of dogs, were better correlated with the decreases in RBC HCN levels than with decreases in plasma HCN concentrations \(\text{(table 3)}\).

A role for the red blood cell in the detoxication of HCN is also compatible with the observation that injected thiosulfate limits the tissue distribution of HCN.\(^{17}\) In fact, a number of isolated reports have claimed demonstration of the enzyme rhodanese in red blood cell.\(^{26-28}\) There is more evidence, however, for the presence of 3-mercaptopropionate:cyanide sulfurtransferase \(\text{(EC.2.8.1.2)}\), which transfers sulfur from 3-mercaptopropionate \(\text{(a metabolite of the amino acid cysteine)}\) to sulfate and cyanide to produce thiosulfate and thiocyate, respectively.\(^{19,29}\)

\[
\text{HSCH}_2\text{COOCO}^- + \text{ENZ} \rightarrow \text{CH}_3\text{COOCO}^- + \text{ENZ}\text{S}\text{SO}_3\text{pyruvate}
\]

\[
\text{ENZ} + \text{SO}_3^- \rightarrow \text{ENZ} + \text{S}_2\text{O}_3^- \text{sulfite thiosulfate}
\]

\[
\text{ENZ} + \text{CN}^- \rightarrow \text{ENZ} + \text{SCN}^- \text{cyanide thiocyanate}
\]

It has been suggested that at high thiosulfate concentration, equation \((2)\) may be reversed.\(^{29}\) The enzyme then could act like rhodanese, as in equation \((3)\). Indeed, electromechanical studies have shown that the rhodanese activity of human erythrocytes is identified with 3-mercaptopropionate sulfurtransferase.\(^{29}\)

This theoretic concept certainly accords with our findings that sodium thiosulfate provides a highly effective, rapid onset antidote to HCN poisoning induced by SNP overdose in dogs. Although previous work has indicated that HCN detoxication is more rapid in the dog,\(^{25}\) there is no reason to believe that the basic mechanism is any different in humans and that our conclusions may not be extrapolated to humans. In practice, it is highly unlikely that any patient would be infused with a dose/dose rate of SNP as high as that given to our dogs and yet, in all of the animals given thiosulfate, HCN clearance and correction of metabolic disturbances were attained rapidly. It therefore is our opinion that thiosulfate does not need to be given as a simultaneous infusion each time SNP is used. Such a practice would result in many patients unnecessarily receiving an extra drug with the use of additional or special infusion routes and equipment. Bolus injection is simpler, allows selection, and, as we have demonstrated, is very effective.

As already indicated, a very brief period of hypotension was the only observed side effect of the injection of sodium thiosulfate. This is particularly notable, since the drug, at a mean concentration of 64%, was given as a rapidly injected bolus into the pulmonary artery. It is normally recommended that a 25% solution of thiosulfate should be given by slow intravenous injection at a rate of 2.5 to 5 ml/min.\(^{30}\) In fact, thiosulfate is an extremely safe HCN antagonist.\(^{31}\) In this it may be contrasted with cobalt EDTA and nitriles. The former is toxic in high dose,\(^{32,33}\) while the nitriles have the distinct disadvantages of 1) reducing the effective haemoglobin concentration by 25-40%,\(^{30,31}\) due to methaemoglobin formation; and 2) exerting a hypertensive action.\(^{33}\) The rationale for the use of nitriles lies in the allegedly low capacity and slow rate of thiosulfate-sulfur transfer to HCN, the combination of HCN with methaemoglobin being a more rapid reaction producing a synergistic effect with thiosulfate.\(^{31}\) Our work has demonstrated that, in fact, injected thiosulfate alone encourages the rapid formation of SCN from HCN. While it is true that this study does not fully reflect the effects of acute cyanide ingestion, the quantities of HCN involved are comparable and the dose used therefore must be very nearly a lethal one \(\text{(see above)}\). Despite this, thiosulfate was capable of dealing with the HCN released. We therefore question whether nitriles, with their inherent drawbacks, are necessary in the treatment of cyanide poisoning from any cause, particularly when additional supportive measures can play a large part in recovery from cyanide poisoning.\(^{34}\) In any case, the use of nitriles certainly would seem to be superfluous when dealing with HCN intoxication from SNP infusion.

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