Pharmacologic EEG Suppression during Cardiopulmonary Bypass: Cerebral Hemodynamic and Metabolic Effects of Thiopental or Isoflurane during Hypothermia and Normothermia


We have determined the effects of thiopental or isoflurane upon cerebral blood flow (CBF) and the cerebral metabolic rate for oxygen (CMRO₂) when these agents are used in sufficient dose to attain a deep burst suppression pattern on the electroencephalogram (EEG) during hypothermic and normothermic cardiopulmonary bypass (CPB). Thirty-one patients undergoing coronary artery bypass graft surgery were anesthetized with fentanyl 0.1 mg·kg⁻¹, and were randomly allocated to one of three groups: control (no further anesthetics during bypass and continuous EEG activity), thiopental treatment (EEG suppression), or isoflurane treatment (EEG suppression). Hypothermia (25–29°C) was routinely induced at onset of nonpulsatile cardiopulmonary bypass. In the treatment groups, thiopental or isoflurane were used during bypass to achieve a deep burst suppression pattern. Cerebral blood flow and cerebral metabolic rate for oxygen were determined during hypothermia and upon rewarmin to normothermia (37°C). Pharmacologic EEG suppression with either isoflurane or thiopental was associated with lower cerebral metabolic rate than control values during both hypothermic and normothermic bypass. However, only thiopental-induced EEG suppression was associated with lower cerebral blood flow than control. Cerebral blood flow during isoflurane-induced EEG suppression was in control to values in spite of the reduced cerebral metabolic rate. (Key words: Anesthesia: cardiovascular. Anesthetics, intravenous: thiopental. Anesthetics, volatile: isoflurane. Brain: blood flow; oxygen consumption. Hypothermia: induced. Monitoring: electroencephalography. Surgery: cardiac.)

In a recent clinical study of the neuropsychiatric sequelae of open heart surgery under normothermic cardiopulmonary bypass (CPB), persistent deficits were found in 7.5% of a control group, while patients subjected to pharmacologic EEG suppression with thiopental during CPB had no persistent deficits.1 This report provided the first evidence of cerebral protection by a barbiturate in humans, and its efficacy was attributed to the reduction of cerebral metabolic rate for oxygen, though this was not measured. The mean dose of thiopental required to maintain EEG suppression throughout CPB was 39.5 mg·kg⁻¹, and the cost of cerebral protection was myocardial depression necessitating increased frequency of administration of inotropic agents, prolonged anesthesia, prolonged time to extubation, and somnolence for up to 3 days. Though it has been stated that this therapeutic modality is now indicated for patients undergoing open chamber heart surgery, the authors urged that... other drugs equally effective in producing EEG suppression without the hemodynamic consequences and persistence of thiopental should be sought.11 Hypothermia is commonly used during CPB to increase the tolerance of the brain to ischemia by reducing the cerebral metabolic rate, and experimental studies have shown that isoflurane, like the barbiturates, reduces cerebral metabolism in a dose-dependent fashion with maximal effect achieved when EEG suppression occurs, and with no direct toxic effect on cerebral metabolic pathways.4,5 Though isoflurane is not without hemodynamic effects, these are dose-dependent and non-persistent. As a major physiologic determinant of cerebral protection is considered the reversible reduction of cerebral oxygen consumption at the time of reduced oxygen delivery, our study assessed the cerebral hemodynamic and metabolic effects of pharmacologic EEG suppression with thiopental and compared them to those occurring with isoflurane during hypothermic and normothermic CPB.

Methods

The study was approved by the Institutional Committee on Human Research, and written informed consent to their participation was obtained from patients scheduled for coronary artery bypass graft surgery. Patients with evidence of cerebrovascular disease or valvular heart disease were excluded from this study. Prior to surgery, patients were randomly allocated to control, thiopental, or isoflurane groups.

After premedication with sublingual lorazepam 0.06 mg·kg⁻¹ and intramuscular morphine sulphate 0.15 mg·kg⁻¹, patients were brought to the operating room where ten EEG leads were affixed in a standard parasagittal bipolar block montage. The EEG was moni-
tored and recorded continuously throughout surgery with a sensitivity of 7 μV and a pass band filter of 0.5–70 Hz. A radial arterial cannula and peripheral venous cannula were inserted under local anesthesia. A central venous or pulmonary arterial catheter was introduced via the right internal jugular vein, either before induction of anesthesia (under infiltration block) or after induction of anesthesia. The same internal jugular vein was then punctured percutaneously, and a 15 cm 16 Fr catheter was passed in a cephalad direction to the jugular bulb to allow sampling of cerebral venous blood. Routine postoperative radiographs were examined to confirm correct placement of these catheters.

Anesthesia was induced and maintained with fentanyl 0.1 mg·kg⁻¹, using succinylcholine or pancuronium bromide to facilitate tracheal intubation. Further doses of pancuronium bromide were used to maintain muscle relaxation. The lungs were ventilated using a Bain non-rebreathing system with a fresh gas flow of 60 ml·kg⁻¹·min⁻¹ of oxygen. Ten scintillation counters were positioned around the head, five over each hemisphere, and connected to a Novo Cerebrograph 10a⁶ to permit determination of cerebral blood flow (CBF) at five periods during surgery. The first measurements were made following sternotomy prior to cannulation for CPB (pre-CPB), and the last measurements were made following protamine administration after CPB (post-CPB), by intravenous injection of ¹³³Xe 5–10 mCi in 6 ml 0.9% saline. Expired air was continuously sampled from the endotracheal tube to obtain end-tidal Xenon concentration used subsequently to correct for isotope recirculation.⁶ Measurements were also made after 15 and 30 min of hypothermic CPB (cold CPB₁₅, cold CPB₃₀; temperature 24.9–29.0 °C), and after rewarming during normothermic CPB (warm CPB; temperature 37.0 °C), by injection of the isotope into the arterial port of the pump oxygenator. In preliminary studies, a scintillation detector placed over the aortic inflow cannula during CPB showed less than 1% recirculation of the original xenon bolus; therefore, a correction for recirculation was not applied. Regional cerebral blood flow (rCBF) under each counter was determined by stochastic (height/area) analysis of the ¹³³Xe washout curve over 15 min.⁷ The calculated flow values were corrected for changes in the xenon partition coefficient resulting from alterations in hemoglobin concentration and temperature.⁵ Mean hemisphere blood flow was calculated from the “hemispheric clearance curve” constructed by summing the outputs of the five detectors on each side.

Blood gas analysis was performed on arterial and jugular venous blood samples at 37 °C and corrected to the patients nasopharyngeal temperature by the factors of Kelman and Nunn¹⁰ for determination of oxygen content according to the formula C₄O₂ = (P₄O₂ x s) + (Hb x 1.34 x S₄O₂), where C₄O₂ = oxygen content of sample (either arterial or venous), P₄O₂ = oxygen tension, s = the solubility coefficient of oxygen in blood adjusted for temperature (s = 0.00395 ml·dl⁻¹·mmHg⁻¹ at 26–28 °C, 0.00317 ml·dl⁻¹·mmHg⁻¹ at 36–38 °C), Hb = hemoglobin concentration, and S₄O₂ = oxygen saturation. The cerebral metabolic rate for oxygen (CMRO₂) was calculated as the product of the mean right hemispheric CBF and the arterio-jugular oxygen content difference. Hemodynamic parameters were recorded during the first 5 min of CBF determination. Cerebral perfusion pressure (CPP) was calculated as the mean arterial pressure – mean jugular venous pressure.

Prior to aortic cannulation patients were given heparin 3–4 mg·kg⁻¹, and subsequent doses as necessary to maintain the activated coagulation time (ACT) beyond 400 s. Bypass technique included the use of a membrane oxygenator without arterial line filter and non-pulsatile pump flows of 2–2.5 l·m⁻²·min⁻¹. The perfusionist attempted to achieve normal arterial pH and Pco₂ as measured at 37 °C without correction for patient temperature.¹¹,¹²

Patients allocated to the control group received no additional anesthetic agents. Patients allocated to thiopental treatment group received intravenous doses of thiopental 8 mg/kg¹³, plus further doses sufficient to maintain a predominantly isoelectric burst/suppression EEG pattern during CPB, such that the duration of the isoelectric periods always exceeded the bursts (fig. 1). The first three patients received thiopental throughout CPB to an empirically predetermined maximum of 24 mg·kg⁻¹, but, as EEG suppression for the full period of CPB could not be consistently maintained within this total dose limit, further patients received thiopental only for the first 15 min of hypothermic CPB (cold CPB₁₅) and upon rewarming to normothermia (warm CPB). In these patients, continuous EEG activity was allowed to resume in the interval between measurement periods cold CPB₁₅ and warm CPB, and no measurement was attempted at cold CPB₃₀. Patients allocated to isoflurane treatment group received isoflurane, in air and oxygen, via the pump oxygenator in sufficient concentration (read from the vapouriser dial setting) to attain a predominantly isoelectric burst suppression EEG pattern (fig. 1).

Mean arterial pressure during CPB was kept above 40 mmHg by administration of phenylephrine as required. Hypertension (pressure greater than 90 mmHg) was treated with chlorpromazine (maximum dose 25 mg) or sodium nitroprusside. Immediately before termination of CPB, 0.5–1.0 g calcium chloride was injected via the pump for all patients. After termination of CPB, fluids and inotropes were used as necessary to achieve satisfac-
tory hemodynamic parameters. Protamine was administered in sufficient dosage to restore the ACT to pre-CPB values.

Data were submitted to one-way analysis of variance (ANOVA), and Scheffes test for multiple comparisons was used to identify differences when ANOVA was significant to $P < 0.05$. For nonparametric data, chi-square testing using Bonferroni's correction for multiple comparisons, with $P < 0.05/k$, was used to determine significance.

Results

As shown in table 1, there were no significant differences in demographics between groups. Subjects were predominantly male, and the age range was 45–73 yr.

Routine postoperative radiography showed no malposition of the jugular catheters. Nasopharyngeal temperature, arterial carbon dioxide tension, hemoglobin concentration, cerebral perfusion pressure, and mean arterial pressure in each group at each stage of the study are shown in table 2. One hundred and twenty-six acceptable CBF measurements were made in 31 patients (table 2). There were no significant variations in rCBF and intrahemispheric CBF values were comparable. Right hemispheric CBF was utilized for determination of CMRO₂ to conform with the side of the jugular bulb catheter.

Compared to either the thiopental group or the control group, phenylephrine was required significantly more often ($P < 0.0005$) when isoflurane was used to suppress the EEG, whereas administration of vasodilators was required more frequently ($P < 0.0005$) in the control group versus the other groups.

Continuous low frequency EEG activity was recorded throughout CPB in control group patients (fig. 1). During hypothermic CPB (temperature range 24.9–29°C), 1.1 ± 0.3% isoflurane produced EEG burst suppression, and, during normothermic CPB (temperature range 36.5–38.1°C), 2.4 ± 0.4% isoflurane was required to maintain EEG burst suppression. In the thiopental group, the mean total dose of barbiturate needed to achieve 30–40 min EEG burst suppression was 17 ± 4 mg/kg.

There were no significant differences in pre-CPB values of CBF and CMRO₂ (tables 3, 4; figs. 2, 3) EEG

<table>
<thead>
<tr>
<th>Table 1. Patient Demographics</th>
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<tr>
<td></td>
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<tr>
<td>Sex M/F (n)</td>
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<tr>
<td>Age (yr)</td>
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<tr>
<td>Weight (kg)</td>
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<tr>
<td>Duration of CPB (min)</td>
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</table>

Results are mean ± SD. Results N.S. at $P < 0.05$. CPB = cardiopulmonary bypass.
TABLE 2. Intraoperative Variables

<table>
<thead>
<tr>
<th>Data Measurements (n)</th>
<th>Pre-CPB</th>
<th>Cold CPB&lt;sub&gt;15&lt;/sub&gt;</th>
<th>Cold CPB&lt;sub&gt;50&lt;/sub&gt;</th>
<th>Warm CPB</th>
<th>Post-CPB</th>
</tr>
</thead>
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<tr>
<td>Control</td>
<td>11</td>
<td>11</td>
<td>6</td>
<td>11</td>
<td>10</td>
</tr>
<tr>
<td>Thiopental</td>
<td>7</td>
<td>7</td>
<td>2</td>
<td>7</td>
<td>6</td>
</tr>
<tr>
<td>Isoflurane</td>
<td>10</td>
<td>11</td>
<td>8</td>
<td>9</td>
<td>10</td>
</tr>
<tr>
<td>Temperature (°C)</td>
<td></td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>Control</td>
<td>35.5 ± 0.6</td>
<td>26.8 ± 1.0</td>
<td>26.8 ± 1.2</td>
<td>37.1 ± 0.4</td>
<td>35.8 ± 0.3</td>
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<tr>
<td>Thiopental</td>
<td>35.0 ± 0.6</td>
<td>26.5 ± 1.4</td>
<td>(25.0, 25.2)</td>
<td>37.1 ± 0.3</td>
<td>35.8 ± 0.3</td>
</tr>
<tr>
<td>Isoflurane</td>
<td>35.7 ± 0.3</td>
<td>26.6 ± 0.9</td>
<td>28.3 ± 0.6†</td>
<td>37.3 ± 0.5</td>
<td>35.6 ± 0.6</td>
</tr>
<tr>
<td>PaCO&lt;sub&gt;2&lt;/sub&gt; (mmHg)</td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>Control</td>
<td>36 ± 5</td>
<td>41 ± 3</td>
<td>37 ± 3</td>
<td>34 ± 5</td>
<td>37 ± 3</td>
</tr>
<tr>
<td>Thiopental</td>
<td>35 ± 1</td>
<td>42 ± 1</td>
<td>(26, 44)</td>
<td>32 ± 6</td>
<td>37 ± 3</td>
</tr>
<tr>
<td>Isoflurane</td>
<td>36 ± 2</td>
<td>42 ± 4</td>
<td>42 ± 8</td>
<td>36 ± 2</td>
<td>37 ± 3</td>
</tr>
<tr>
<td>MAP (mmHg)</td>
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<td></td>
<td></td>
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<tr>
<td>Control</td>
<td>95 ± 11</td>
<td>72 ± 14</td>
<td>83 ± 5</td>
<td>60 ± 13</td>
<td>77 ± 13</td>
</tr>
<tr>
<td>Thiopental</td>
<td>91 ± 13</td>
<td>54 ± 12*</td>
<td>(61, 61)</td>
<td>58 ± 10</td>
<td>74 ± 2</td>
</tr>
<tr>
<td>Isoflurane</td>
<td>89 ± 13</td>
<td>52 ± 8*</td>
<td>59 ± 10*</td>
<td>52 ± 4</td>
<td>73 ± 7</td>
</tr>
<tr>
<td>CPP (mmHg)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Control</td>
<td>84 ± 16</td>
<td>66 ± 19</td>
<td>74 ± 8</td>
<td>52 ± 12</td>
<td>67 ± 12</td>
</tr>
<tr>
<td>Thiopental</td>
<td>81 ± 13</td>
<td>44 ± 12*</td>
<td>50 ± 5</td>
<td>48 ± 11</td>
<td>61 ± 5</td>
</tr>
<tr>
<td>Isoflurane</td>
<td>75 ± 11</td>
<td>40 ± 9*</td>
<td>50 ± 11</td>
<td>38 ± 3*</td>
<td>58 ± 7</td>
</tr>
<tr>
<td>Hb (mg/dl)</td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>Control</td>
<td>12.3 ± 0.86</td>
<td>8.6 ± 1.0</td>
<td>8.5 ± 0.7</td>
<td>8.4 ± 1.1</td>
<td>9.1 ± 1.0</td>
</tr>
<tr>
<td>Thiopental</td>
<td>11.8 ± 1.4</td>
<td>7.6 ± 1.0</td>
<td>(7.2, 6.9)</td>
<td>7.7 ± 0.9</td>
<td>8.4 ± 0.9</td>
</tr>
<tr>
<td>Isoflurane</td>
<td>12.4 ± 0.9</td>
<td>8.4 ± 1.0</td>
<td>8.5 ± 0.7</td>
<td>8.4 ± 0.5</td>
<td>8.4 ± 0.4</td>
</tr>
</tbody>
</table>

PaCO<sub>2</sub> at 37°C; MAP = mean arterial pressure; CPP = cerebral perfusion pressure; Hb = hemoglobin concentration; CBF = cerebral blood flow; CPB = cardiopulmonary bypass. Results are mean ± SD.

* P < 0.05 vs. control; †P < 0.05 vs. thiopental.

Pharmacologic EEG suppression with isoflurane during hypothermic CPB (cold CPB<sub>15</sub>) was associated with significantly lower CBF and CMRO<sub>2</sub> than control. Only two CBF measurements were obtained at cold CPB<sub>50</sub> in the thiopental group, and these data points are presented in Table 3, but were not subjected to statistical analysis. Thiopental-induced EEG suppression was also associated with significantly lower CBF and CMRO<sub>2</sub> than control during warm CPB.

Pharmacologic EEG suppression with isoflurane was associated with a significantly lower CMRO<sub>2</sub> than control at each stage of CPB. One patient in the isoflurane group had elevated CBF before CPB (48 ml · 100 g<sup>-1</sup> · min<sup>-1</sup>) and during CPB (27 ml · 100 g<sup>-1</sup> · min<sup>-1</sup> at cold CPB<sub>15</sub>; 24 ml · 100 g<sup>-1</sup> · min<sup>-1</sup> at cold CPB<sub>50</sub>) with similar CMRO<sub>2</sub> to the other patients in the group, and so was considered an outlier; the values are included from statistical analysis. CBF during isoflurane-induced EEG suppression at cold CPB<sub>15</sub> was intermediate between control and thiopental groups, but was not statistically different from either. At cold CPB<sub>50</sub> CBF during isoflurane-induced EEG suppression was again somewhat lower than control, but did not reach statistical significance. During warm CPB, CBF during isoflurane-induced EEG suppression was similar to control and greater than thiopental.

Post-CPB measurements were made 15–30 min after termination of CPB, and in EEG suppression groups, after discontinuation of the agent used with return to continuous EEG activity. CBF and CMRO<sub>2</sub> values after CPB were similar in all three groups.

In the early post-bypass period, ephedrine or an infusion of inotropic agents were used to increase cardiac contractility in 2/12 control patients, 2/7 patients who had been treated with thiopental, and 2/12 patients who had been treated with isoflurane. A further two patients in the control group and two patients who had received thiopental required mechanical assist devices for left ventricular failure attributable to perioperative myocardial ischemia.

TABLE 3. Cerebral Blood Flow (ml · 100 g<sup>-1</sup> · min<sup>-1</sup>)

<table>
<thead>
<tr>
<th></th>
<th>Pre-CPB</th>
<th>Cold CPB&lt;sub&gt;15&lt;/sub&gt;</th>
<th>Cold CPB&lt;sub&gt;50&lt;/sub&gt;</th>
<th>Warm CPB</th>
<th>Post-CPB</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>26.9 ± 5.6</td>
<td>14.6 ± 5.5</td>
<td>12.8 ± 4.1</td>
<td>19.5 ± 4.8</td>
<td>25.4 ± 6.0</td>
</tr>
<tr>
<td>Thiopental</td>
<td>32.4 ± 6.7</td>
<td>8.2 ± 2.5*</td>
<td>(7.2, 12.1)</td>
<td>14.9 ± 1.0†</td>
<td>21.8 ± 3.7</td>
</tr>
<tr>
<td>Isoflurane</td>
<td>24.6 ± 5.8</td>
<td>11.8 ± 2.5</td>
<td>9.2 ± 1.5</td>
<td>19.6 ± 4.7</td>
<td>24.8 ± 3.6</td>
</tr>
</tbody>
</table>

CPB = cardiopulmonary bypass. Results are mean ± SD.

* P < 0.05 vs. control; †P < 0.05 vs. other groups.
TABLE 4. Cerebral Metabolic Rate for Oxygen (ml·100 g⁻¹·min⁻¹)

<table>
<thead>
<tr>
<th></th>
<th>Pre-CBP</th>
<th>Cold CPB&lt;sub&gt;19&lt;/sub&gt;</th>
<th>Cold CPB&lt;sub&gt;21&lt;/sub&gt;</th>
<th>Warm CPB</th>
<th>Post-CBP</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>1.80 ± 0.32</td>
<td>0.41 ± 0.08</td>
<td>0.48 ± 0.11</td>
<td>1.16 ± 0.21</td>
<td>1.38 ± 0.37</td>
</tr>
<tr>
<td>Thiopental</td>
<td>2.56 ± 0.48</td>
<td>0.27 ± 0.02*</td>
<td>0.23 ± 0.37</td>
<td>0.92 ± 0.14*</td>
<td>1.33 ± 0.13</td>
</tr>
<tr>
<td>Isoflurane</td>
<td>1.78 ± 0.41</td>
<td>0.29 ± 0.04*</td>
<td>0.30 ± 0.06*</td>
<td>0.76 ± 0.16*</td>
<td>1.17 ± 0.24</td>
</tr>
</tbody>
</table>

CPB = cardiopulmonary bypass. Results are mean ± SD.

* P < 0.05 vs. control.

Discussion

This is the first report of the cerebral metabolic and hemodynamic effects of thiopental or isoflurane-induced EEG suppression during CPB in humans.

The CBF measurement techniques used in the present study (intravenous and intra-aortic injections) will both yield a slight underestimate of "true" CBF due to the presence of Xenon in the extracerebral (scalp) tissues. Estimates of grey matter blood flow obtained by bicompartamental analysis are not significantly affected by this extracerebral contamination, but estimates of white matter and weighted mean blood flows are underestimated by 10–15%.

The use of noncompartmental stochastic analysis (height/area) for calculating mean CBF is less sensitive to this contamination, and integration to 15 min rather than infinity further reduces the effect of the extracerebral compartment. The stochastic method produces estimates of mean CBF which are within 2–5% of the "true" mean flow over a wide range of flow rates.

For determination of CMRO₂, sampling of effluent cerebral venous blood from a jugular bulb catheter is representative of the venous drainage from all brain structures due to mixing in the confluence of the venous sinuses. Although there is some drainage of extracerebral tissue via the cortical emissary veins, blood sampled from the jugular bulb is contaminated to less than 3% by extracerebral flow.

Patients undergoing CPB for closed heart surgery were selected as subjects for this study, as they are less at risk of cerebral embolism which might cause pathologic derangements of CBF, complicating the interpretation of the data. The radioxenon washout technique has previously been used to measure CBF in humans during cardiac surgery in an attempt to elucidate the effects of cardiopulmonary bypass on the cerebral circulation, and postoperatively to ascertain the incidence of diffuse cerebral injury.

Many factors influence CBF and CMRO₂, and it has been reported that CBF is independent of mean arterial pressure over the range 30–110 mmHg using the described bypass technique, implying that cerebral autoregulation is maintained. The ventilation parameters used in this study produced mild hypocapnia before and after CPB. Our intention during CPB was to achieve normocapnia on arterial blood gas analysis measured at 37°C and uncorrected for patient temperature, and values achieved were comparable between groups at each period (table 2).

The profile of intraoperative CBF and CMRO₂ changes seen in the control group for this study is similar to that previously reported from this unit, in which the anesthetic technique varied primarily in the premedication used. The Pre-CBP CBF and CMRO₂ values are lower than normal values for the awake patient, and are attributable to anesthesia, mild hypothermia, and hypocapnia (table 2). The Pre-CBP CBF values are comparable to those obtained by Henriksen et al. using the Initial Slope Index method following intravenous injection of radio-Xenon. Goyier and Prough measured CBF in humans during hypothermic CPB and found low values comparable to the control group presented here. Both this and our previous study have documented the low CMRO₂ attained during hypothermic CPB in humans, and we postulate that CBF/CMRO₂ coupling is responsible for the low CBF. By contrast, Henriksen et al. reported cerebral hyperperfusion during hypothermic CPB, but their bypass technique included the addition of CO₂ to maintain normal temperature-corrected arterial P<sub>CO₂</sub> values. The effects of acid/base management on CBF and CMRO₂ during hypothermic CPB are discussed by us elsewhere.

Fig. 2. CBF values in control, thiopental, and isoflurane groups; see text for discussion. Values are mean ± SD. O = oulying result excluded from statistical analysis (patient in isoflurane group); ○ = individual values for CBF in thiopental group at CPB<sub>19</sub> (n = 2); CBF = cerebral blood flow; CPB = cardiopulmonary bypass. *P < 0.05 vs. control.
EEG suppression was achieved in all patients treated with either thiopental or isoflurane. The initial dose of thiopental (8 mg·kg⁻¹) was determined from a report by Quasha et al. noting its efficacy in inducing prolonged burst suppression during hypothermic CPB. Isoflurane has similarly been shown to be an effective EEG suppressant during hypothermic CPB by Loomis et al. They reported onset of burst suppression at a higher average isoflurane vaporizer setting (2.2%), which, exclusive of vaporizer performance, may partly reflect the lower dosage of narcotic their patients had received (average fentanyl 0.07 mg·kg⁻¹). In addition, their primary end-point was reduction of MAP rather than induction of EEG burst suppression. We have reported isoflurane-induced EEG suppression during non-cardiac surgery occurring at vaporizer settings similar to those required during normothermic CPB.

CMRO₂ at burst-suppression was similar with either agent during hypothermia (approximately 0.3 ml·100 g⁻¹·min⁻¹ at 28°C), and presumably reflects the basal cerebral oxygen consumption at those temperatures. The proportionate reduction in CMRO₂ achieved by pharmacologic EEG suppression from the anesthetized, continuous-EEG control state at either temperature was of the order of 30%, and is in good agreement with the findings of Astrup et al. who used pentobarbital or lidocaine to induce EEG suppression in a canine laboratory model of hypothermic and normothermia CPB. Our results also support the hypothesis of Steen et al. that thiopental only reduces CMRO₂ during CPB by reducing functional electrical activity, and is only additive with hypothermia in the presence of EEG activity. For example, it is evident from our data that hypothermia to 26–30°C, which does not extinguish EEG activity, provides more cerebral metabolic depression than does pharmacologic EEG suppression at 37°C (table 4). We are not aware of any laboratory studies of isoflurane effect on cerebral metabolism during hypothermic CPB, but Newberg et al. have demonstrated that isoflurane, like thiopental, only reduces CMRO₂ by reducing functional cerebral activity, and that no further reduction in CMRO₂ occurs when the EEG becomes isoelectric.

In the thiopental group, CBF was reduced in conjunction with CMRO₂. Reductions in CBF would be expected to produce a proportionate reduction in the delivery of emboli to the cerebral circulation. The cerebroprotective effects of thiopental reported by Nussmeier et al. may, therefore, be primarily due to cerebral vasoconstriction rather than a primary reduction in CMRO₂. A reduction of CBF was not achieved in the isoflurane group. There was a tendency to lower CBF associated with isoflurane-induced EEG suppression during hypothermic CPB, but it did not attain statistical significance in this study. A biphasic effect of isoflurane on CBF has been noted in baboons; at less than 1 MAC isoflurane, CBF is reduced in conjunction with CMRO₂, but, at greater than 1 MAC isoflurane, cerebral vasodilatation occurs and CBF returns to baseline levels, despite further reduction in CMRO₂. This uncoupling of the CBF/CMRO₂ relationship has been documented in humans during isoflurane-induced hypotension and isoflurane-induced EEG suppression. There may, therefore, be a real reduction of CBF during EEG suppression with 1% isoflurane during hypothermic CPB which, because of sample size, this study has failed to confirm statistically. EEG suppression with 2.5–3.5% isoflurane during normothermic CPB is undoubtedly associated with a degree of cerebral hyperperfusion relative to the reduced CMRO₂. One possible mechanism for this vasodilatation is an increase in brain cyclic-AMP.

A potentially limiting side effect of barbiturate-induced EEG suppression is known to be persistent depression of myocardial contractility. Upon reviewing our preliminary results, we found that three of 22 patients studied up to that point had evidence of severe ischemic left ventricular failure after CPB. Two of them had received thiopental. Though there was no evidence that thiopental was responsible for myocardial ischemia, we were concerned that the barbiturate might reduce the response to inotropic drugs, and felt that we were not justified in continuing to use thiopental in association with our high-dose fentanyl anesthetic technique for patients with ischemic heart disease, as they are not the primary risk group for embolic cerebral complications. At completion of our study, one further patient (in the control group) had sustained severe ischemic left ventricular failure.

In summary, we have demonstrated that EEG suppression with thiopental during CPB in humans reduces both CBF and CMRO₂. Isoflurane is also effective in inducing EEG suppression and reducing CMRO₂ dur-
ing CPB, although it does not reduce CBF during nor-
moothermic CPB when high concentrations are required to
maintain EEG suppression. These differences in ef-
efect on CBF may be important in considerations of poten-
tial cerebral protection against cerebral emboli
global or focal ischemia associated with cardiopulmo-
nary bypass. Hypothermia is a potent metabolic depres-
sant and, in the presence of EEG activity, its effects can
be augmented by pharmacologic EEG suppression.
Clinical outcome studies are indicated to assess the rela-
tive benefits of hypothermia and pharmacologic EEG
suppression in patients undergoing open or closed heart
surgery during CPB.

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