A Closed Xenon Anesthesia Delivery System


XENON was discovered in 1898 and first used as an anesthetic in 1951.1 It is more potent than nitrous oxide (N\textsubscript{2}O) with a minimum alveolar concentration (MAC) of approximately 71%, and its low blood-gas partition coefficient of 0.14 (recently recalculated as 0.115) results in rapid induction and recovery.2–7 Interest in xenon is high because of medical and environmental concerns about N\textsubscript{2}O; however, use may be constrained high cost.8–10 Fully closed rebreathing circuits are one method of minimizing costs. It is possible to close a conventional circle system, with the anesthesiologist varying the fresh gas flow to match uptake via the lungs. However, better control might be achieved by automation of this procedure.8–11 We describe a closed breathing system that may have advantages versus other designs.

Materials and Methods

Breathing System

The breathing system comprised a disposable carbon dioxide absorber–valve unit (King Systems Corp., Noblesville, IN) plus hoses; a mechanical ventilator (Kontron ABT 4000, Milan, Italy) and a bellows system modified by one of the authors (J. D.) to allow oxygen entry into the circle (fig. 1). At equilibrium, oxygen was added automatically at end-inspiration to keep the volume constant, replacing gas uptake from the closed circle. The system volume was approximately 2,170 ml (770 ml circle volume + 900 ml functional residual capacity + 500 ml tidal volume delivered).

With a xenon–oxygen mixture in the circle, this mechanical arrangement would result in a gradual increase in circle oxygen concentration as uptake of both gases was replaced with oxygen. This tendency was offset by delivery of xenon boluses to the circle via a computer-controlled magnetic valve (Airmatic-Allied Inc., Wilmington, OH), to maintain a stable measured oxygen concentration. The computer control algorithm was written using Labview software (National Instruments, Austin, TX) and based on the algorithm of Luttropp et al.8 It was a model-based, forward-feedback loop-control system in which a volume of xenon was delivered to an estimated distribution volume to achieve a target measured oxygen concentration by dilution. The estimated distribution volume comprised the volumes of the breathing system and lungs plus a volume representing the continual loss of gas by tissue uptake. Initial xenon overdose to the circle while the algorithm optimized an appropriate distribution volume was prevented by use of a deliberately low 2 l starting value. The computer controlled and recorded xenon bolus volumes by varying the magnetic valve opening time; the flow rate through the valve was constant and known. The condition for xenon delivery was oxygen concentration greater than target value and bellows nearly empty (oxygen about to be added) and previous delivery at 60 s or more (to allow mixing of previous xenon bolus). When running closed, the bellows never filled completely, and, thus, no gas was spilled. Xenon was delivered to the expiratory limb to maximize mixing before it entered the lungs. Leak testing was performed via ventilation of a dummy lung. Absence of movement of the oxygen flow indicator confirmed a lack of volume loss. A small leak across the automatic oxygen addition valve mechanism could theoretically be tolerated as any leaked gas would be flushed back by oxygen during the following cycle.

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Anesthesia and Surgical Preparation

Experiments were performed in five purebred, large, white male pigs. All complied with the 1986 Animals (Scientific Procedures) Act of the United Kingdom in accordance with the National Institutes of Health guidelines on the use of experimental animals. Anesthesia was induced using a halothane–oxygen–nitrous oxide mixture, and the trachea intubated with use of an uncuffed 6.5-mm ID cuffed tube (SIMS Portex Ltd., Hythe, UK) without neuromuscular blockade. After right external jugular vein cannulation, anesthesia was maintained with an alphaxalone–alphadolone infusion at 15 mg·kg\textsuperscript{-1}·h\textsuperscript{-1}, and mechanical ventilation...
continued with use of a 30% oxygen–70% nitrogen mixture.

**Measurements**

The volume of each xenon bolus ($V_1$) stored in the computer database was given by the equation:

\[
V_1 = \text{xenon flow through valve (l/s)} \times \text{duration of valve opening (s)}
\]

Summation of all values of $V_1$ gives the total xenon volume used at ambient temperature and pressure as estimated by the computer ($V_c$). Weighing of the xenon cylinder before and after each experiment allowed a more accurate calculation of the total volume used ($V_w$). Small errors in the values of $V_1$ as a result of the opening and closing times of the magnetic valve could be reduced by multiplying each by a correction factor defined as $V_w/V_c$.

A Servomex 570A paramagnetic oxygen analyzer (Servomex Ltd., Crowborough, Sussex, UK) was used to measure the circle oxygen concentration and data was recorded and continuously fed to the control algorithm. Xenon concentration was simultaneously measured using a calibrated Minison ultrasonic analyzer (Thomas Swan and Co., Ltd., Cambridge, UK). The sample gas was dried via passage through a 10-ml container of silica gel desiccant beads upstream of the analyzers, to reduce xenon measurement error, and returned to the circle otherwise unchanged.

**Experimental Protocol**

A standardized 30-min nitrogen washout maneuver was performed using a Mapleson D system and a mechanical ventilator with a fresh gas flow of 10 l/min, set at 14 breaths/min, and a tidal volume varied to maintain a constant partial pressure of arterial carbon dioxide ($P_{aCO_2}$). The closed breathing system, which was primed...
with oxygen, then was connected. The oxygen concentration was reduced to 80, 60, 40, and then 30%, with two cycles of the algorithm permitted at each of these steps for the algorithm to optimize the estimated xenon distribution volume (1 min/cycle). The final 30% oxygen target remained unaltered for the remainder of the experiment, which lasted 2 h in five animals but was extended to a total of 4 h in one animal. Nitrogen concentration was measured intermittently using a mass spectrometer (model SX200; VG Quadrupoles Ltd., Middlewich, Chesire, UK). Values are expressed as the median and range.

Results

The weight of the pigs was 33 kg (range, 32–39 kg). The median volume of xenon used for the 2-h administration periods (at 22°C and atmospheric pressure) was 7.9 l (range, 7.4–9.1 l). In the 4-h administration, for a 33-kg pig, this volume was 13.1 l (table 1). More than half the xenon necessary for 2 h administration was used in the first 30 min.

An example of the performance of the system during a 2-h period is shown graphically in figure 2.

By the 90- to 120-min measurement interval, the median xenon volume necessary decreased to 1 l every 30 min, and in the pig that received xenon for 4 h, xenon expenditure continued at a similar rate.

Each time a new lower target oxygen concentration was set, the xenon concentration increased in a stepwise fashion, stabilizing within three cycles of the algorithm and within 5 min. The median nitrogen concentration in the breathing system after 2 h was 9.4% (range, 9–11.6%) and was 14.5% after 4 h.

Discussion

The system functioned as intended. We performed a computer simulation using the NARKUP 4.11 program (Drs. D. C. White and G. G. Lockwood, Northwick Park Hospital, Harrow, UK), which uses a xenon blood-gas partition coefficient of 0.14. The simulated xenon expenditure for our system was 4.9 l after 2 h. The observed median xenon expenditure of 7.9 l after 2 h is good compared to the "ideal" value and is similar to the 7.6 l observed by Lutropp et al., who used a functionally closed exchanger system in pigs of similar weight. In engineering terms, our breathing system should be closed, with zero gas loss; however, this is difficult to achieve in practice. Our dynamic leak test relied on a rotating vane to show leaks. These have inertia and friction; therefore, a slight leak might be missed. A static leak test also would have been advisable. Xenon diffuses well through certain materials, for example, silicone rubber. Lutropp et al. showed a loss of 1.5 l via silicone hoses over 2 h in similar experiments. In the current study, the hoses, the gas sampling, and the return lines were made of polyethylene, through which xenon dif-

<table>
<thead>
<tr>
<th>O₂ concentration (%) in circle</th>
<th>100, decreasing to 30</th>
<th>30</th>
<th>30</th>
<th>30</th>
<th>30</th>
</tr>
</thead>
<tbody>
<tr>
<td>Time interval (min)</td>
<td>0–15</td>
<td>15–30</td>
<td>30–60</td>
<td>60–90</td>
<td>90–120</td>
</tr>
<tr>
<td>Median Xe expenditure over time interval (l)</td>
<td>4.1</td>
<td>0.8</td>
<td>1.5</td>
<td>0.9</td>
<td>1.0</td>
</tr>
<tr>
<td>Range</td>
<td>3.0–4.4</td>
<td>0.5–1.5</td>
<td>1.2–1.6</td>
<td>0.7–1.3</td>
<td>0.7–1.1</td>
</tr>
<tr>
<td>Median Xe expenditure over time interval (ml/kg)</td>
<td>121</td>
<td>24</td>
<td>41</td>
<td>28</td>
<td>28</td>
</tr>
<tr>
<td>Range</td>
<td>94–133</td>
<td>13–47</td>
<td>35–48</td>
<td>18–39</td>
<td>21–33</td>
</tr>
</tbody>
</table>

Xe = xenon; O₂ = oxygen.

Table 1. Xenon Expenditure

Fig. 2. Example of a 2-h xenon administration period in pig 4. Target oxygen concentration was set to 80, 60, 40, and then 30%. The measured oxygen concentration in the breathing system and the cumulative xenon expenditure are shown.
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References