Coiling of Stimulating Perineural Catheters

To the Editor.—We read with great interest Dr. Boezaart’s excellent review on continuous perineural catheters and would like to comment on the optimal length of catheter to be threaded beyond the needle tip.

While Dr. Boezaart advocates inserting the catheter 5–10 cm past the needle tip for nonstimulating (blind) catheters and 3–5 cm for stimulating catheters, we would like to point out that the optimal length of catheter to be introduced beyond the needle tip still remains speculative. With blind catheters, some authors have inserted as little as 3 cm, whereas others have threaded as much as 20 cm. The rationale behind introducing such a great length of catheter seems to rest for the most part on the belief that the perineural sheath can be dilated with a small amount of saline or local anesthetic to permit subsequent threading of a catheter toward the plexic roots. However, studies on femoral catheters reveal that only a minority (23–40%) achieve a position close to the lumbar plexus roots: In fact, most tend to course medially in the direction of the psoas muscle or laterally in the direction of the iliacus muscle. It is perhaps this unpredictability in (blind) catheter migration that is responsible for the 40% secondary analgesic block failure rate.

Stimulating perineural catheters, by permitting real-time observation of muscular twitches, allow the operator to manipulate both needle and catheter until the desired length of catheter has been threaded alongside the nerve. Therefore, it should theoretically be easier to direct a stimulating a catheter toward the plexic roots. Thus, in our practice (which uses exclusively stimulating catheters), we anchor the body of the catheter forms a coil that is illustrated by figure 2: The body of the catheter forms a coil that is situated outside the dye-delineated perineural sheath. Therefore, only a fraction (the tip) of the catheter introduced beyond the needle is actually situated inside the nerve’s sheath. This may explain the “all or nothing” phenomenon often seen with perineural catheterization: When the right combination of needle shaft angulation and bevel orientation has been found, the catheter can be easily and swiftly fed through the needle while preserving the same amplitude of motor stimulation. In all likelihood, the tip has been anchored and remains stationary while its shaft is being inserted (and looped) through the needle.

Although it can be argued that coils may contribute to anchor and stabilize perineural catheters subcutaneously, they could also in theory predispose to knotting during removal of the catheter. In the literature, cases of knotting have been reported after advancement of femoral and fascia iliaca catheters 10–20 cm beyond the needle tip. Coiling was thought to be the underlying mechanism. We have previously reported the case of a retained (and surgically extracted) stimulating infracavicular catheter that was inserted only 4 cm beyond the needle tip. The risk of ensnaring the nerve is another possible complication of looping. In our series, despite a relatively high rate of catheter dislodgement (10.5%), no kinking or knotting was noted. All catheters were removed uneventfully. Therefore, despite the high rate of catheter dislodgement (10.5%) reported by a recent series of 1,416 patients, cooling cannot be used as a means of catheter stabilization. Subcutaneous catheter tunneling may offer a safer alternative.

In summary, although the optimal distance of stimulating catheter to be threaded beyond the needle tip remains speculative, Dr. Boezaart’s

Table 1. Distance and Minimal Stimulating Threshold for Noncoiled and Coiled Catheters

<table>
<thead>
<tr>
<th>Distance, cm</th>
<th>Noncoiled</th>
<th>Coiled</th>
<th>P Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>2.5</td>
<td>0.07 ± 0.21</td>
<td>0.51 ± 0.20</td>
<td>0.22</td>
</tr>
<tr>
<td>3</td>
<td>0.07 ± 0.21</td>
<td>0.51 ± 0.20</td>
<td>0.22</td>
</tr>
<tr>
<td>4</td>
<td>0.07 ± 0.21</td>
<td>0.51 ± 0.20</td>
<td>0.22</td>
</tr>
<tr>
<td>5</td>
<td>0.07 ± 0.21</td>
<td>0.51 ± 0.20</td>
<td>0.22</td>
</tr>
<tr>
<td>6</td>
<td>0.07 ± 0.21</td>
<td>0.51 ± 0.20</td>
<td>0.22</td>
</tr>
<tr>
<td>7</td>
<td>0.07 ± 0.21</td>
<td>0.51 ± 0.20</td>
<td>0.22</td>
</tr>
<tr>
<td>8</td>
<td>0.07 ± 0.21</td>
<td>0.51 ± 0.20</td>
<td>0.22</td>
</tr>
</tbody>
</table>

Data are mean ± SD.

Illustrated by figure 2: The body of the catheter forms a coil that is situated outside the dye-delineated perineural sheath. Therefore, only a fraction (the tip) of the catheter introduced beyond the needle is actually situated inside the nerve’s sheath. This may explain the “all or nothing” phenomenon often seen with perineural catheterization: When the right combination of needle shaft angulation and bevel orientation has been found, the catheter can be easily and swiftly fed through the needle while preserving the same amplitude of motor stimulation. In all likelihood, the tip has been anchored and remains stationary while its shaft is being inserted (and looped) through the needle.

In summary, although the optimal distance of stimulating catheter to be threaded beyond the needle tip remains speculative, Dr. Boezaart’s

![Distribution of coiled and noncoiled catheters](http://anesthesiology.pubs.asahq.org/pdfaccess.ashx?url=/data/journals/jasa/931069/)

Fig. 1. Distribution of coiled and noncoiled (straight) catheters according to the distance in centimeters threaded beyond the needle tip.

David C. Warltier, M.D., Ph.D., served as Handling Editor for this exchange.
Ultrasound-guided Perineural Catheters Have Already Arrived

Certainly, when discussing perineural local anesthetic infusions, ultrasound deserves a little more respect.

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2. Ilfeld BM, Morey TE, Enneking K: Continuous infraclavicular brachial plexus block for postoperative pain control at home. Anesthesiology 2002; 96:1297–304
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To the Editor:—I read with much dismay the article by Boezaart regarding perineural infusion of local anesthetics. As a practicing anesthesiologist at a teaching hospital, it was concerning to see the flagship journal of our specialty present a review of perineural catheter placement with such little regard for emerging technology. Many who practice regional anesthesia on a daily basis consider ultrasound to be the most significant advance in several decades. Boezaart, however, dismisses ultrasound as a technique that “works well for superficial nerves (when it is not really needed) . . . [but] is not sufficient to identify deeper nerves, especially in very obese patients (where it is most needed).” The author goes on to say that ultrasound is “not likely to replace nerve stimulation for continuous nerve block.”

I would encourage the author to review a number of quality publications by Marhofer et al., Chan et al., and Sandhu and Capal to name a few, which demonstrate the superiority of ultrasound techniques when compared with nerve stimulation. At the University of Utah (Salt Lake City, Utah), we no longer use or teach nerve stimulator techniques. More than 2 yr ago, our techniques were completely transitioned to ultrasound guidance. Our residents perform more than 3,000 blocks per year (more than 1,000 indwelling catheters) using only ultrasound guidance, with satisfying results both for the resident and for the patient.

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To the Editor.—Dr. Boezaart’s review article titled “Perineural Infusion of Local Anesthetics”1 represents a commendable attempt to educate the reader on a complex and evolving topic. In reading this article, we were surprised by the author’s use of a study performed in volunteers to support the claim that the use of a stimulating catheter was associated with “around” 100% success compared with 65–85% for a nonstimulating technique. Studies conducted in patients certainly did not support this concept.2 It is unfortunate that Dr. Boezaart did not acknowledge the value of continuous lumbar plexus blocks for hip replacement.3-5 We were surprised by the author’s use of a study performed in volunteers close to the tourniquet or surgical field, electromyographic and conduction nerve studies cannot allow such a differentiation, and by default, the anesthesiologists is usually blamed. In our practice, we favor the use of approaches away from the tourniquet and the site of the surgery.6

The problem of continuous nerve blocks and anticoagulation is undoubtedly an important issue for regional anesthesiologists. In his article, the author referred to recommendations made by the American Society of Regional Anesthesiologists, but after going to the American Society of Regional Anesthesiologists Web site, we could not verify the source of these recommendations. Moreover, the authors of this letter have several thousand patient experiences with the combination of continuous lumbar plexus and thromboprophylaxis using various anticoagulants (aspirin, warfarin, low-molecular-weight heparin, and fondaparinex). As long as these anticoagulants are administered for the prevention of deep vein thrombosis and pulmonary embolism and not for their treatment, we found no reason to take any precaution when placing or removing the lumbar plexus catheter.7 For total knee arthroplasty,8,9 we would also like to stress the value of a continuous sciatic nerve block for postoperative analgesia. In our experience, 80% of patients undergoing total knee replacement report sciatic pain. Finally, although the author recommended the use of 0.2% ropivacaine for perineural infusion, he also recommend the use of 0.5% bupivacaine for the initial injection. Bupivacaine is recognized to be more toxic than ropivacaine. Therefore, it is surprising that the author would continue to recommend its use for the initial bolus injection. Certainly, in our practice, we have eliminated the use of bupivacaine.

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8. Graf BM, Abraham I, Eberbach N, Kunst G, Stowe DF, Martin E: Differences in cardiotoxicity of bupivacaine and ropivacaine are the result of physicochemical and stereoselective properties. ANESTHESIOLOGY 2002; 96:1427–34

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In Reply.—I thank Dr. De Tran et al. for sharing their experience and insights. With regard to their criticism: I did not advocate “inserting the catheter 5–10 cm past the needle tip for nonstimulating . . . catheters.” Because I have little personal experience with nonstimulating catheters, I was merely reflecting what the authors who I cited typically do. I agree that the optimal distance of catheter advancement is speculative at this stage of our knowledge, but the catheter that I typically use (StimuCath; Arrow International, Reading, PA) is an armored catheter, which is floppy when the stylet is removed. The stylet goes down the catheter almost all the way but ends 5 cm from its tip. That, plus my experience of not having any problems with 3–to-5-cm catheter advancement, has led me to believe that 3–5 cm is probably appropriate. I am eagerly awaiting the results of the study planned by Dr. De Tran and colleagues in this regard.

The belief that the perineural sheath can be dilated with small amounts of saline has not yet been validated by research. In fact, we tried to demonstrate this on the perineural tissue of anesthetized pigs under direct vision and could only show that the tissue became edematous and the nerve stimulation was lost, making stimulation via the catheter useless. However, if practitioners still subscribe to this belief, I suggest that they use 5% dextrose in water to “open up the space” so that nerve stimulation is still possible.1 Research is ongoing to evaluate this notion.

I share the author’s sentiments regarding stimulating catheters. It would be interesting to know for what blocks the 81 catheters of which 70% were found to be coiled up were placed. Based on my own experience, I share the belief that most catheters coil up, but like Dr. De Tran et al., I do not have any objective data for my belief. It does seem that close to 100% of paravertebral catheters coil up, whereas a relatively small number of femoral catheters do. I am anxiously awaiting the results of their studies. I do believe, however, that the coiling is of no consequence as long as it does not involve too much of the catheter (advanced more than 5 cm), and its tip, where the drug comes out, is in the same fascia compartment as the nerve. This may be a problem with multiorifice catheters, because most of the drug comes out the most proximal orifice.

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Ensuring a nerve in the coil is at this stage only a theoretical possibility because, to the best of my knowledge, no such case has yet been reported in the literature.

We tunnel all catheters, and I agree with Dr. De Tran that it should probably be done routinely.

I share Dr. Swenson’s enthusiasm for ultrasound as a promising “emerging technology” to aid in peripheral nerve blocks. I did not have any access to the unpublished results of Dr. Swenson and his students, and I am eagerly awaiting their publication. Surely, my omission of these results cannot be regarded as a flaw in my article.

I have reread the excellent articles by Marhofer et al., Chan et al., and Sandhu and Capal (Dr. Swenson’s references 2–6), and I was again struck by the finding that all five of these reports were on single-injection nerve blocks. Not unlike the rest of the literature, they do not address continuous nerve blocks by ultrasound guidance. I would respectfully remind Dr. Swenson that the title of my article was “Peripheral Injection of Local Anesthetics” and not “Peripheral Injection of Local Anesthetics.”

Based on the current state of our knowledge, it seems clear that for nerves that are “deep” (and it is not uncommon for us to do subgluteal sciatic nerve blocks where the nerve is 15 cm or deeper, although around 8 cm is the rule), the ultrasound technology simply does not help us much. I therefore believe that my statement that ultrasound helps us the most when the nerve is superficial, and the least when the nerve is deep, is factually correct. The experts cited by Dr. Swenson all restricted their studies to superficial nerve blocks.

With the equipment currently available to us, one needs at least three educated hands to place a catheter with ultrasound guidance: one to hold and maneuver the ultrasound probe, one to hold and maneuver the needle, and one to hold and maneuver the catheter. It is hoped that the emerging technology will address this problem soon.

Most ultrasound experts that I discussed this with (some cited above) agree that a combined technique of finding the nerve with the ultrasound and needle and then placing the catheter with a nerve stimulator is optimal at this stage of our development. Most would not rigidly transform to using only ultrasound guidance for catheters at this stage.

Finally, one of the experts cited by Dr. Swenson has as recently as July 2006 written, “Unfortunately ultrasound cannot help to guide the catheter into the sheath compartment. Because catheters generally curl up as they are advanced, multiple cross sectional views of the catheter are captured on ultrasound; thus the position of the catheter tip cannot be determined accurately.”

With nerve stimulation, there is no problem in accurately determining the position of the catheter tip.

Drs. Chelly and Casati raised some interesting issues. The worst technique for continuous nerve block will have exactly the same outcome as the best technique, in terms of postoperative pain, if not all of the nerves that innervate that joint are blocked and the patients also receive effective multimodal analgesia. The studies that demonstrated no difference between stimulating and nonstimulating catheters were all performed on femoral nerve blocks for knee1 and hip1 surgery, ignoring the other nerves that innervate the knee and hip joints. Similarly, for example, the best femoral nerve blocks for knee3 and hip4 surgery, ignoring the other nerves that innervate that joint are blocked and the patients also receive effective multimodal analgesia. The studies that demonstrated no difference between stimulating and nonstimulating catheters improve effective placement? A randomized, controlled, and observer-blinded trial. Anesth Analg 2005; 100:1503–10

I agree with the sentiments of Drs. Chelly and Casati regarding anticoagulation and continuous peripheral nerve blocks. Unfortunately, when the consensus guidelines were formulated by the American Society of Regional Anesthetologists task force, they stated, “The Consensus Statements on Neuraxial Anesthesia and Anticoagulation may be applied to plexus and peripheral techniques.” An important factor that stimulated the development of continuous nerve and plexus blocks was precisely the growing popularity of low-molecular-weight heparin, and I am eagerly awaiting the publication of the “several thousand patient experiences” referred to by Drs. Chelly and Casati to strengthen the overdue case against this statement in the American Society of Regional Anesthetologists consensus document.

If even the sentiments of Drs. Chelly and Casati that 80% of patients need a sciatic block after total knee arthroplasty were correct (in my experience it is closer to 20%), it would mean that 20% of Drs. Chelly and Casati’s patients (80% of my patients) would receive a nerve block that they do not need. I therefore offer our patients postoperative sciatic nerve blocks if they need it. That would mean that 100% of the patients who received the sciatic blocks actually needed them. Furthermore, my experience is that single-injection sciatic nerve blocks are long-lasting blocks as opposed to the posterior knee pain of total knee arthroplasty, and surgeons get nervous if the inevitable “foot drop” persists.

Although I agree that ropivacaine is a better choice than bupivacaine, bupivacaine is much cheaper than ropivacaine and quite suitable when patients cannot afford the more expensive drug.

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References


(Accepted for publication August 22, 2006.)
To the Editor—In the context of the current debate around the deleterious effects of the cyclooxygenase-2 inhibitors in patients with increased risks for cardiovascular diseases, we read with great interest the study published by Nussmeier et al.1

The authors have studied, as intended, a population that was certainly at low risk for cardiovascular disease, and, indeed, the number of cardiovascular events was small and identical in both groups (five events per group [table 3 from the original article1]).

In the discussion section, the authors highlight the fact that the size of the studied groups is too small to show any statistically significant difference in the number of cardiovascular events, because the study was only powered to show a difference in a combined multisystem endpoint.

Nevertheless, we believe that readers should not be left with an impression of possibly false security and conclude that the administration of a cyclooxygenase-2 inhibitor in a low-risk population does not increase the occurrence of cardiovascular events. This has simply not been demonstrated in this study, partly because of two methodologic problems that should be addressed and have not been discussed in the article.

First, the analyses were performed on a modified intent-to-treat population, were the patients who did not receive the treatment were excluded from the analysis after randomization (four patients in the placebo group and eight patients in the parecoxib–valdecoxib group). The creation of a “modified intent-to-treat population” could lead to the destruction of the similarity of the groups after randomization. This problem is further increased by the fact that the reasons for not administering the treatment are not given in the article (as per the CONSORT statement2) and that twice as many patients were excluded from the analysis after randomization (four patients in the placebo group and two in the parecoxib–valdecoxib group).

In both groups, patients were lost to follow-up (five in the parecoxib–valdecoxib group and two in the placebo group). Although this is never clearly stated in the article, the usual definition of lost to follow-up implies that the primary endpoint (the combined incidence of postrandomization adverse events) is not known.

We computed the maximal possible bias to judge the robustness of the statistical results. In this method, all of the missing patients (eight excluded from analysis and five lost to follow-up) in the parecoxib–valdecoxib group are considered to have presented a cardiovascular event, in contrast to none of four excluded and two lost to follow-up in the placebo group. This is done with inclusion of all of the randomized patients. The computation of the risk ratio for presenting a cardiovascular event is given in table 1.

In contrast, when none of the 13 patients in the parecoxib–valdecoxib group are considered to have presented a cardiovascular event, but the 6 patients in the placebo group are considered to have presented a cardiovascular event, the results are given in table 2.

We concentrated this analysis on the cardiovascular adverse events because they are currently the most feared and highlighted adverse events of the cyclooxygenase-2 inhibitors.

One can see from these results that, with the data from the article, it is impossible to eliminate, from a statistical point of view, that the parecoxib–valdecoxib combination does increase the frequency of cardiovascular events in a low-risk population. But one can easily eliminate, with a high probability, that the placebo administration increased the frequency of cardiovascular events.

This should be clearly stated in the discussion section, where we have the impression that more emphasis is given to the demonstration of analgesic efficacy, which was a secondary endpoint, than to the lack of demonstration within a certain statistical probability of the primary endpoint, which was clearly stated to be the safety.

The second problem of this article is the marginal statistical power of the study. The authors state that “The sample size of 500 patients per treatment arm provided at least 80% power to detect a doubling of the 4% estimated background incidence of all predefined adverse events combined.” In fact, to provide this sort of power, you need at least 550 patients per treatment with the primary endpoint available. With the available data in the study, calculated on 525 patients per arm, you have, at best, a power of 74%, which leaves 26% of chance (or more exactly bad luck) to face a type II error of not being able to detect a difference of 4% that would really exist. This lack of power is further aggravated by the fact that the true incidence of the combined adverse event was even lower than expected, and demonstrating a doubling of the 3.2% event rate in the placebo group would necessitate 697 patients per treatment to maintain an 80% power. All of these power calculations are true if we suppose that the authors accepted a 5% risk of type I error, which in fact was never stated in the article.

The conclusion about the cardiovascular safety in a low-risk population, even after this study, is that we do not know with any certainty. And we should not be afraid to say so.

Table 1. Risk Ratio Computation Considering That All Missing Patients from the Parecoxib–Valdecoxib Group Have Presented a Cardiovascular Event, in Contrast to None in the Placebo Group

<table>
<thead>
<tr>
<th>Adverse Event</th>
<th>Placebo (n = 529)</th>
<th>Parecoxib–Valdecoxib (n = 533)</th>
<th>Risk Ratio (95% CI)</th>
<th>P Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cardiovascular event (%)</td>
<td>5 (0.9%)</td>
<td>18 (3.4%)</td>
<td>3.57 (1.34–9.55)</td>
<td>0.011</td>
</tr>
</tbody>
</table>

CI = confidence interval.

Table 2. Risk Ratio Computation Considering That All Missing Patients from the Placebo Group Have Presented a Cardiovascular Event, in Contrast to None in the Parecoxib–Valdecoxib Group

<table>
<thead>
<tr>
<th>Adverse Event</th>
<th>Placebo (n = 529)</th>
<th>Parecoxib–Valdecoxib (n = 533)</th>
<th>Risk Ratio (95% CI)</th>
<th>P Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cardiovascular event (%)</td>
<td>11 (2.1%)</td>
<td>5 (0.9%)</td>
<td>0.45 (0.16–1.29)</td>
<td>0.137</td>
</tr>
</tbody>
</table>

CI = confidence interval.
Edgard Engelman, M.D.,* Jean-Corentin Salengros, M.D.
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In Reply—We wish to thank Drs. Engelman and Salengros for their comments. They correctly state that our analyses were performed on a modified intent-to-treat population, in which the patients who did not receive the treatment were excluded from analysis after randomization. The following paragraph of the International Conference on Harmonisation guidance on the use of statistics in clinical trials1 makes it clear that the practice of removing patients who do not receive treatment is not without merit:

In some situations, it may be reasonable to eliminate from the set of all randomized subjects any subject who took no trial mediation. The intention-to-treat principle would be preserved despite the exclusion of these patients provided, for example, that the decision of whether or not to begin treatment could not be influenced by knowledge of the assigned treatment. In other situations it may be necessary to eliminate from the set of all randomized subjects any subject without data post randomization. No analysis should be considered complete unless the potential biases arising from these specific exclusions, or any others, are addressed.

Regarding the current study,2 it should be kept in mind that we are discussing only 12 patients who did not receive treatment (8 in the parecoxib–valdecoxib group and 4 in the placebo group) in a study that included more than 1,050 patients—approximately 1% of the randomized population. It is highly unlikely that the treatment responses of these few patients would have differed from the responses of the patients included in the analysis.

Drs. Engelman and Salengros also state that ‘this problem is further increased by the fact that the reasons for not administering the treatment are not given in the article . . . and that twice as many patients were excluded from the parecoxib–valdecoxib group as from the placebo group.’ However, given that there were 12 patients, the fact that 8 were assigned to one treatment and 4 to the other is not at all unlikely; in fact, it has more than a 19% probability of occurring by chance alone. It should also be understood that the number of patients not treated could just as easily have been 8 placebo and 4 parecoxib–valdecoxib patients. If one used this distribution in the sensitivity analysis suggested by the authors, one would conclude that parecoxib is actually cardioprotective. This scenario shows that the suggested analysis is inappropriate.

Drs. Engelman and Salengros then compute the risk ratio for a cardiovascular event if those patients in the parecoxib–valdecoxib intent-to-treat group who never received the treatment (n = 8) plus the 5 patients lost to follow-up in the parecoxib–valdecoxib group had all experienced a cardiovascular event. This sensitivity analysis is based on the premise that the patients who were not treated or lost to follow-up could have had vastly greater risk than the patients included in the analysis. That is, although the percentage of patients having a cardiovascular event in both placebo and parecoxib–valdecoxib groups was actually 1%, this sensitivity analysis assumes that 100% of the excluded patients in the parecoxib–valdecoxib group could have had an event, which is not even remotely consistent with the observed data. In fact, we have gone back to look at the raw data from patients who did not receive treatment or were lost to follow-up and determined that none of these in either group experienced any cardiovascular thromboembolic events during the limited time that they were under observation.

Drs. Engelman and Salengros also state that they concentrated this analysis on the cardiovascular adverse events and could easily eliminate the possibility that placebo administration increased the frequency of cardiovascular events. This is simply not true, as shown by the confidence intervals and P values produced by these two extreme analyses. Again, the results would change dramatically with a slight change in the number of missing data points in the placebo group.

Drs. Engelman and Salengros also mention that the risk of a type I error was never stated in the article. Certainly, we agree that this information should have been included. However, the considerable breadth of the confidence interval of the risk ratio (0.3–3.5) that we reported2 makes it highly unlikely that increasing the power of the study would reveal a significant difference in cardiovascular event rates between the treatment and placebo groups. In addition, we have already acknowledged this limitation in the Discussion, in which we state:

Another limitation of the study is the sample size. Although this was the largest trial of any nonsteroidal antiinflammatory drug in patients undergoing noncardiac surgery, the number of adverse events was relatively small and possibly inadequate to detect a particular safety signal. This is especially true for cardiovascular thromboembolic events, given the low level of risk in this population compared with CABG [coronary artery bypass graft] surgery patients. Nevertheless, this population was representative of the majority of patients who undergo major surgery.

In summary, we appreciate Drs. Engelman and Salengros’ thoughtful comments and speculations. Nonetheless, our interpretation of the results of our study—the largest such clinical trial yet performed worldwide—and the conclusions we draw from them remain unchanged.

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**Ultrasound Guidance for Peripheral Venous Access: A Simplified Seldinger Technique**

To the Editor—Occasionally, patients for ambulatory surgery present with difficult peripheral venous access. Because only short-term access is needed, I prefer to avoid central venous cannulation and the associated risks. In these cases, I have begun using ultrasound guidance and a simplified Seldinger technique for upper extremity peripheral venous access. A description of the technique and the results of a series of 10 procedures follows.

In this technique, a tourniquet is applied to the upper arm. Alcohol or povidone iodine is used to prepare the antecubital fossa and distal upper arm. Disposable, nonsterile gloves are worn, and draping is unnecessary. A 10-5 MHz ultrasound probe (Sonosite Titan, L38; Sonosite, Bothell, WA) is covered with gel, an occlusive dressing (3M Tegaderm, St. Paul, MN), and then additional sterile gel. Using a transverse view, an appropriate, nonthrombosed vein is identified. This may be the cephalic, basilic, or brachial vein. The brachial artery and median nerve are identified, so they can be avoided. Lidocaine is infiltrated 1-2 cm distal to the planned insertion site. I then use a spring-wire guide/catheter over needle assembly, usually used for arterial catheterization (Arrow arterial catheterization set FA-0420; Arrow International, Reading, PA). It consists of a 10.8-cm, 20-gauge catheter over a 22-gauge thin wall needle with an integral 0.46-mm-diameter spring wire guide. Using a transverse or longitudinal view and a free hand technique, the needle is inserted into the selected vein under real-time imaging. When ultrasound imaging shows the needle in the vein and venous blood flashback appears, I ask an assistant to advance the spring wire guide. Alternately, I drop the ultrasound probe and advance the wire myself. Next, the catheter is advanced over the wire, the tourniquet is released, and the needle/wire assembly is removed.

Table 1 shows the results of a series of 10 consecutive ultrasound-guided peripheral venous access procedures performed by the author. Patients were included in this series if at least two attempts at standard peripheral venous access failed or no adequate sized veins were visible or palpable in the upper extremity. Success was defined by the ultrasound view of the catheter in the vein and free flow of venous appearing blood from the catheter. Each separate skin puncture was considered an attempt. Time was defined as the interval from initial skin puncture until success was achieved or the procedure was aborted. Nine of 10 catheter insertions were successful. There was an average of 1.3 attempts per procedure. The time required for a procedure averaged 140 s.

Ultrasound guidance is used commonly for peripherally inserted central catheters and occasionally for standard peripheral venous access. In emergency medicine applications, Keyes et al.1 found that a 5-cm catheter over needle infiltrated 8% of the time, and therefore, a longer catheter might be useful for these deeper veins. Sandhu and Sidhu2 recommended long needle-mounted catheters or a Seldinger technique for deep veins. Using a guide wire may help to ensure that an advancing catheter enters the vein properly. However, a standard Seldinger technique takes more time and requires sterile gloves, draping, and a table to hold the wire, tissue dilator, and catheter.

Ultrasound guidance for peripheral venous access may be improved by the use of a simplified Seldinger technique. The Arrow catheter set is commonly used for arterial catheterization, so anesthesiologists should be familiar with its use. The series described here demonstrates that this technique has a good success rate and the procedure takes a relatively short time to complete. Ultrasound guidance using this catheter may prevent multiple puncture attempts and decrease the use of unnecessary central venous catheters.

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References


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Table 1. Series of 10 Ultrasound-guided Peripheral Venous Access Procedures

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<thead>
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<th>Vein</th>
<th>Success</th>
<th>Attempts</th>
<th>Time, s</th>
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<tr>
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<tr>
<td>Basilic</td>
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<td>2</td>
<td>483</td>
</tr>
<tr>
<td>Brachial</td>
<td>Yes</td>
<td>1</td>
<td>65</td>
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<tr>
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<td>2</td>
<td>201</td>
</tr>
<tr>
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<td>1</td>
<td>62</td>
</tr>
<tr>
<td>Basilic</td>
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</tr>
<tr>
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<tr>
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<td>48</td>
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<tr>
<td>Mean average</td>
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</table>

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