Mild Endotoxemia during Mechanical Ventilation Produces Spatially Heterogeneous Pulmonary Neutrophilic Inflammation in Sheep

Eduardo L. V. Costa, M.D., Ph.D.,* Guido Musch, M.D.,† Tilo Winkler, Ph.D.,‡ Tobias Schroeder, Ph.D.,§ R. Scott Harris, M.D.,∥ Hazel A. Jones, Ph.D.,# Jose G. Venegas, Ph.D.,** Marcos F. Vidal Melo, M.D., Ph.D.***

ABSTRACT

Background: There is limited information on the regional inflammatory effects of mechanical ventilation and endotoxemia on the production of acute lung injury. Measurement of 18F-fluorodeoxyglucose (18F-FDG) uptake with positron emission tomography allows for the regional, in vivo, and noninvasive assessment of neutrophilic inflammation. The authors tested whether mild endotoxemia combined with large tidal volume mechanical ventilation bounded by pressures within clinically acceptable limits could yield measurable and anatomically localized neutrophilic inflammation.

Methods: Sheep were mechanically ventilated with plateau pressures = 30–32 cm H2O and positive end-expiratory pressure = 0 for 2 h. Six sheep received intravenous endotoxin (10 ng·kg⁻¹·min⁻¹), whereas six did not (controls), in sequentially performed studies. The authors imaged with positron emission tomography the intrapulmonary kinetics of infused 13N-nitrogen and 18F-FDG to compute regional perfusion and 18F-FDG uptake. Transmission scans were used to assess aeration.

Results: Mean gas fraction and perfusion distribution were similar between groups. In contrast, a significant increase in 18F-FDG uptake was observed in all lung regions of the endotoxin group. In this group, 18F-FDG uptake in the middle and dorsal regions was significantly larger than that in the ventral regions. Multivariate analysis showed that the 18F-FDG uptake was associated with regional aeration (P < 0.01) and perfusion (P < 0.01).

Conclusions: Mild short-term endotoxemia in the presence of heterogeneous lung aeration and mechanical ventilation with pressures within clinically acceptable limits produces marked spatially heterogeneous increases in pulmonary neutrophilic inflammation. The dependence of inflammation on aeration and perfusion suggests a multifactorial basis for that finding. 18F-FDG uptake may be a sensitive marker of pulmonary neutrophilic inflammation in the studied conditions.

What We Already Know about This Topic
❖ Endotoxemia causes pulmonary inflammation, but whether this is homogeneous throughout the lungs during large tidal volume ventilation is not known

What This Article Tells Us That Is New
❖ In sheep with mild endotoxemia and large tidal volume ventilation within ranges used clinically, neutrophil inflammation occurred heterogeneously, related to local aeration and perfusion
❖ In this situation, regional pulmonary injury might occur by increased mechanical stress and localized inflammation

ENDOTOXEMIA and mechanical ventilation are frequently associated in clinical practice.1–3 Acute lung injury (ALI) due to endotoxemia has been characterized as a generalized process of lung inflammation.1–6 In contrast, a key element in ALI is the heterogeneous spatial distribution of lung aeration.7–10 Studies to date indicate that endotoxin exposure either preceding or accompanying injurious mechanical ventilation augments lung inflammation and injury.11–13 Indeed, isolated cell, ex vivo and in vivo small animal studies showed that exposure to high tidal volumes and endotoxin leads to increased production of neutrophil-attracting cytokines,11–16 increased neutrophil counts in the bronchoalveolar lavage,13,16 and worsening of gas exchange and respiratory mechanics.12,13,17 Although these investigations illustrate basic inflammatory processes induced by the combination of excessive lung strain and endotoxin, they do not allow for a straightforward translation of those findings to specific regions of the heterogeneously aerated and perfused lungs.
Materials and Methods

The experimental protocols were approved by the Massachusetts General Hospital Subcommittee on Research Animal Care (Boston, Massachusetts). Twelve sheep (22.3 ± 5.9 kg, approximately 3 months old) were fasted overnight and premedicated with intramuscular ketamine (4 mg/kg) and midazolam (2 mg/kg). After intravenous induction of anesthesia with ketamine (4 mg/kg), an endotracheal tube was inserted. General anesthesia was maintained with a continuous infusion of propofol and fentanyl titrated to heart rate and blood pressure. Pancuronium 0.1 mg/kg at induction and repeated every 90 min (0.02–0.04 mg/kg) was used for muscle paralysis. Each sheep was placed supine in the PET scanner (Scanditronix PC4096; General Electric, Milwaukee, WI) with the caudal end of the field of view just superior to the dome of the diaphragm. After a recruitment maneuver, they were mechanically ventilated with $P_{\text{plat}} = 30–32$ cm H$_2$O, $P_{\text{EEP}} = 0$, inspired O$_2$ fraction = 0.3 (adjusted to an arterial O$_2$ saturation >0.88), inspiratory-to-expiratory time ratio = 1:2, respiratory rate = 18 breaths/min or higher to maintain the arterial carbon dioxide pressure (PaCO$_2$) between 32 and 45 mmHg. If PaCO$_2$ were less than 32 mmHg when respiratory rate = 18 breaths/min, a variable dead space was added to the breathing circuit aiming at that PaCO$_2$ range. Physiologic data were collected, and PET scans were acquired both at the start of the protocol and after 2 h of mechanical ventilation, except for the FDG scan performed at the end of the study. After the initial set of scans, six sheep (endotoxin group) received a continuous infusion of propofol and fentanyl titrated to heart rate

PET Imaging Protocol and Processing

The experimental system and methods of analysis have been described in detail.29,30,33–35 Scans consisted of 15 cross-sectional slices of 6.5-mm thickness over a 9.7-cm-long axis field, providing three-dimensional data for an estimated 70% of the total lung volume.35 For each slice, resulting images, consisting of 128 × 128 voxels of 6 × 6 × 6.5-mm size, were low-pass filtered to 12 × 12 mm to a final volumetric resolution of approximately 0.9 cm$^3$.

1. Transmission scans: used to correct for attenuation and to calculate regional gas fraction ($F_{\text{gas}}$). We categorized pulmonary parenchyma as nonaerated ($F_{\text{gas}} <0.1$), poorly aerated (0.1 ≤ $F_{\text{gas}} <0.5$), normally aerated (0.5 ≤ $F_{\text{gas}} <0.85$), and hyperinflated ($F_{\text{gas}} ≥ 0.85$).36,37

2. Emission scans

a. Intravenous $^{13}$N-nitrogen ($^{13}$NN)-saline: used to measure regional pulmonary perfusion and shunt from the lung tracer kinetics after a bolus injection of $^{13}$NN-saline during a 60-s apnea at mean lung volume.35,38 Because of the low solubility of nitrogen in blood and tissues (partition coefficient water-to-air is 0.015 at 37°C), in perfused and aerated regions, vir-
tually all $^{13}$NN diffuses into the alveolar airspace at first pass, accumulating in proportion to regional perfusion. In regions with shunting alveolar units, there is a peak of tracer concentration in the early PET frames, corresponding to arrival of the bolus of tracer with pulmonary blood flow, followed by a decrease toward a plateau due to lack of retention of $^{13}$NN in nonaerated units. The magnitude of this decrease is related to regional shunt. Perfusion and shunt fraction were calculated with a tracer kinetics model.39

b. Intravenous $^{18}$F-FDG: used for quantification of regional neutrophilic inflammation.29,30 After $^{13}$NN clearance, $^{18}$F-FDG (5–10 mCi) was infused at a constant rate through the jugular catheter over 60 s, and, starting at the beginning of $^{18}$F-FDG infusion, sequential PET frames (9 × 10 s, 4 × 15 s, 1 × 30 s, 7 × 60 s, 15 × 120 s, 1 × 300 s, and 3 × 600 s) were acquired while pulmonary arterial blood was sampled at 5.5, 9.5, 25, 37, and 42.5 min. Blood samples (1 ml) were spun down, and the activity of plasma was measured in a gamma counter cross-calibrated with the PET scanner. The plasmatic activities of those samples were used to calibrate the blood-pool region of interest (ROI) (see Selection of Voxels for Analysis section) and obtain an image-derived input function taking into account partial-volume and spillover effects.33 $^{18}$F-FDG PET scans were acquired only after injury because of the 110-min half-life of $^{18}$F-FDG.

Inside cells, $^{18}$F-FDG is phosphorylated by hexokinase to $^{18}$F-FDG-6-phosphate, which accumulates in proportion to cellular metabolic rate. $^{18}$F-FDG net uptake rate ($K_i$), a measure of cellular metabolic activity, was calculated at the ROI level by fitting the $^{18}$F-FDG kinetics with a three-compartment model of Sokoloff et al.39 To account for potential effects of lung inflation and blood volume on regional $K_i$, we standardized $K_i$ by lung tissue, thus computing a specific $K_i$ as $K_i = K_i/F_{\text{tissue}}$, where $F_{\text{tissue}} = (1-F_{\text{gas}}-F_{\text{blood}})$ and $F_{\text{blood}}$ is the fractional volume of the blood compartment obtained from the model of Sokoloff et al. $K_i$ is proportional to $^{18}$F-FDG uptake per gram of lung tissue.

Patlak analysis41 was used to compute $K_i$ at the voxel level and construct parametric images (fig. 1) and illustrations of regional kinetics (fig. 2).

### Selection of Voxels for Analysis

Identification of the aerated lung fields was done by thresholding the transmission scans. Early frames of the $^{13}$NN-saline infusion emissions scans were used to identify nonaerated perfused lung fields. The combination of those fields resulted in the final lung mask for each animal. We manually excluded areas corresponding to main bronchi and large pulmonary vessels.

Three ROIs of same vertical height (ventral, middle, and dorsal) were defined by dividing the three-dimensional lung mask with two horizontal planes and used for quantification of regional $F_{\text{gas}}$, and $^{13}$NN and $^{18}$F-FDG kinetics.

A blood-pool ROI was defined by thresholding the regional activity of $^{13}$NN during the first 5 s after $^{13}$NN-saline injection. During this time, $^{13}$NN is confined mostly to the right heart cavities and pulmonary arteries, and only a minor amount has diffused into the alveolar gas volume.33

### Histologic Analysis

Lungs from four animals of the control group and five animals of the endotoxin group were excised at the end of the experiment and fixed with Trump’s fixative (4% formaldehyde and 1% glutaraldehyde in phosphate-buffered saline) at a pressure of 25 cm H$_2$O. A block of lung tissue was sampled from ventral and dorsal regions and embedded in paraffin. Sections of 5-μm thickness were cut, mounted, and stained with hematoxylin-eosin for light microscopy. Lung neutrophils were counted in 40 randomly selected high-power (400×) fields per animal (10 per region) by two investigators, who were blinded to the group assignment. In addition, perivascular and alveolar edema, alveolar hemorrhage, septal thickening, and capillary congestion were evaluated semiquantitatively with a four-grade scale (absent = 0, mild = 1, moderate = 2, and marked = 3).
Statistical Analysis

Variables were tested for normality with the Shapiro-Wilk test. We expressed values as means and standard deviations for normally distributed variables and median and interquartile ranges (25–75%) otherwise. For normally distributed variables, we used the independent samples Student t test for comparisons between groups and paired t tests for comparisons between time points in the same group. For not normally distributed variables, we used the Wilcoxon rank sum test for comparisons between groups and the Wilcoxon signed rank test for comparisons between time points in the same group. Regional Fgas, perfusion, and shunt in ventral, middle, and dorsal regions within and between groups and before and after mechanical ventilation were compared with a linear mixed-effect model. This model was chosen because the analysis involved observations in the same regions of the endotoxin group, consistent with higher 18F-FDG uptake in the endotoxin than in the control group. (C, D) Quantification of the pulmonary 18F-FDG kinetics as Patlak plots, in which the regional 18F-FDG activity (CROI(t)) normalized to the plasmatic 18F-FDG activity (Cp(t)) is plotted against the integral of plasma activity normalized to plasma activity. In these plots, the slope of the linear regression represents 18F-FDG uptake. Note the different regional 18F-FDG uptakes.
similar considerations as those described earlier. The multivariate regression analysis was applied. All statistical tests were two-tailed, and the significance was set at $P < 0.05$.

**Results**

**Global Physiologic Variables**

By experimental design, PEEP and $P_{\text{plat}}$ were kept at 0 and 30 cm H$_2$O, respectively. Along 2 h of mechanical ventilation, mean tidal volume decreased in both groups, significantly in the endotoxin group (table 1). In this group, respiratory rate was increased to maintain the PaCO$_2$ within the predefined acceptable range. The shift in PaCO$_2$ toward the upper limit of the accepted range was associated with a nonsignificant difference in pH (table 1). Oxygenation was reduced in the endotoxin group during the 2 h of study ($P < 0.02$).

Hemodynamics was stable during the experiment and comparable between groups. Circulating neutrophil counts decreased significantly in the endotoxin group ($P < 0.02$).

**Table 1.** Cardiovascular and Respiratory Variables and Neutrophil Counts in Peripheral Blood at Baseline and after 2 h of Mechanical Ventilation

<table>
<thead>
<tr>
<th></th>
<th>Control (n = 6)</th>
<th>Endotoxin (n = 6)</th>
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</thead>
<tbody>
<tr>
<td></td>
<td>Baseline</td>
<td>2 h</td>
</tr>
<tr>
<td>Mean systemic arterial pressure (mmHg)</td>
<td>74 ± 14</td>
<td>81 ± 13</td>
</tr>
<tr>
<td>Mean pulmonary artery pressure (mmHg)</td>
<td>14 ± 8</td>
<td>15 ± 9</td>
</tr>
<tr>
<td>Cardiac output (l/min)</td>
<td>4.9 ± 2.7</td>
<td>5.2 ± 2.8</td>
</tr>
<tr>
<td>$P_{\text{V}_{2}O_2}$ (mmHg)</td>
<td>47 ± 7</td>
<td>48 ± 7</td>
</tr>
<tr>
<td>$F_{\text{SO}_2}$ (%)</td>
<td>0.32 ± 0.04</td>
<td>0.35 ± 0.05</td>
</tr>
<tr>
<td>Tidal volume (ml/kg)</td>
<td>17.8 (17.0–23.5)</td>
<td>14.9 (13.9–19.0)</td>
</tr>
<tr>
<td>Respiratory rate (bpm)</td>
<td>19 ± 1</td>
<td>19 ± 3</td>
</tr>
<tr>
<td>pH</td>
<td>7.44 ± 0.07</td>
<td>7.41 ± 0.09</td>
</tr>
<tr>
<td>$P_{\text{A}<em>2O_2} / P</em>{\text{FiO}_2}$ (mmHg)</td>
<td>243 ± 66</td>
<td>228 ± 88</td>
</tr>
<tr>
<td>$P_{\text{A}_2CO_2}$ (mmHg)</td>
<td>34 (30–41)</td>
<td>36 (32–51)</td>
</tr>
<tr>
<td>Neutrophil count ($\times 10^{9}/\mu l$)</td>
<td>1.78 (1.63–2.43)</td>
<td>3.89 (3.21–5.72)</td>
</tr>
</tbody>
</table>

Variables are expressed as mean ± SD for normally distributed variables and median and interquartile range (25–75%) otherwise.

*$P < 0.01$ vs. baseline.

$F_{\text{Io}_2}$ = inspired fraction of oxygen; $P_{\text{A}_2CO_2}$ = arterial partial pressure of carbon dioxide; $P_{\text{A}_2O_2} / P_{\text{FiO}_2}$ = ratio between arterial partial pressure of oxygen and $F_{\text{Io}_2}$; $P_{\text{V}_{2}O_2}$ = mixed venous partial pressure of oxygen.

Fig. 3. Gas fraction (A), mean normalized perfusion (B), and shunt fraction (C) in the ventral, middle, and dorsal regions in the control (n = 6) and endotoxin (n = 6) groups of supine sheep at baseline and after 2 h of mechanical ventilation with positive end-expiratory pressure = 0 cm H$_2$O and plateau pressure of 30–32 cm H$_2$O. Data are expressed as box (median, twenty-fifth, and seventy-fifth percentile) and whiskers (range) for each region, group, and time point. On the x axis, “b” indicates measurements at baseline, and “2h” indicates measurements after 2 h of mechanical ventilation. *$P < 0.05$ versus baseline.
Regional Aeration at Baseline and after Mechanical Ventilation

Mean $F_{gas}$ of the imaged lung was similar in both groups at baseline (controls $= 0.57 \pm 0.08$ and endotoxin group $= 0.58 \pm 0.04$, NS) and after 2 h of mechanical ventilation (controls $= 0.56 \pm 0.08$, endotoxin group $= 0.53 \pm 0.06$, NS; figs. 1 and 3). The regional distribution of $F_{gas}$ at baseline and after 2 h of mechanical ventilation was not statistically different between the two groups (fig. 3). $F_{gas}$ decreased significantly after 2 h in the dorsal regions in both groups and in the ventral regions of the control group (fig. 3).

There was no statistical difference in aeration between groups at baseline and after 2 h of mechanical ventilation (fig. 4). Most voxels in the ventral and middle regions were normally aerated, whereas voxels in the dorsal regions were either poorly aerated or nonaerated (fig. 4). The proportion of hyperinflated voxels was small in all regions. After 2 h of mechanical ventilation, the size of the nonaerated compartment increased significantly only in the dorsal ROIs of the endotoxin group ($P = 0.02$).

Perfusion and Shunt Fraction

The regional distribution of perfusion before and at the end of the 2-h period was comparable between groups (figs. 1 and 3). The distribution of perfusion was inversely related with that of aeration along the vertical ROIs. In the ventral and middle regions of both groups, shunt fraction was small and remained unchanged after 2 h of mechanical ventilation (fig. 3). Shunt increased significantly in the dorsal regions of the endotoxin group.

Regional $^{18}$F-FDG Kinetics and Regional Neutrophilic Inflammation

PET assessment of $^{18}$F-FDG kinetics yielded regional plots that were suitable for analysis in all cases of both groups (figs. 2A and B). These plots were characterized by an early peak followed by either a continuously decreasing curve in controls or a plateau or slightly ascending slope in the endotoxin group. Quantification of the kinetics with Patlak plots (figs. 2C and D) or computations derived from the model of Sokoloff et al. (table 2) evidenced the differences in regional $^{18}$F-FDG uptake, particularly high in dorsal ROIs.
In the control group, regional Ki was low, showed a small interanimal variability in all ROIs, and was not significantly different among the ROIs of the same animal (table 2; fig. 5A). Likewise, specific Ki (Kis), representing the standardization of Ki by the amount of lung tissue in the ROI, was not significantly different for the different ROIs (table 2; fig. 5B), despite the differences in regional perfusion and aeration. In contrast, global and regional Ki were larger in the endotoxin group, with mean values of global Ki and Kis more than twofold greater than those measured in the control group (table 2; fig. 5C). Furthermore, there were large differences in Ki and Kis among ROIs in the endotoxin group, in addition to larger interanimal variability in these parameters. On average, Ki and Kis increased progressively from ventral to dorsal ROIs (figs. 5C and D). As a result, Ki in the endotoxin group was 168% and Kis was 46%, larger in dorsal than in ventral ROIs.

Kis increased monotonically with perfusion in the endotoxin group, but not in the controls (figs. 6A and B). A different relationship was observed between Kis and Fgas in the endotoxin group (figs. 6C and D). Kis values in the two extremes of aeration were larger than those at the intermediate Fgas tertile. No changes in Kis were observed in the control

Table 2. 18F-Fluorodeoxyglucose Uptake Rates for the Control and Endotoxin Groups in Ventral, Middle, and Dorsal Lung Regions and in the Whole Lung

<table>
<thead>
<tr>
<th>Region</th>
<th>Control Ki (10^{-3}/min)</th>
<th>Control Kis (10^{-2}/min)</th>
<th>Endotoxin Ki (10^{-3}/min)</th>
<th>Endotoxin Kis (10^{-2}/min)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ventral</td>
<td>4.1 ± 0.4</td>
<td>1.4 ± 0.1</td>
<td>6.0 ± 2.3</td>
<td>2.2 ± 0.7</td>
</tr>
<tr>
<td>Middle</td>
<td>3.7 ± 0.6</td>
<td>1.4 ± 0.2</td>
<td>7.6 ± 3.7</td>
<td>2.9 ± 1.6</td>
</tr>
<tr>
<td>Dorsal</td>
<td>6.5 ± 1.6</td>
<td>1.4 ± 0.2</td>
<td>16.2 ± 7.5</td>
<td>3.1 ± 1.1</td>
</tr>
<tr>
<td>Whole Lung</td>
<td>4.7 ± 0.7</td>
<td>1.6 ± 0.2</td>
<td>10.2 ± 4.4</td>
<td>3.4 ± 1.5</td>
</tr>
</tbody>
</table>

Variables are expressed as mean ± SD.

* P < 0.05 versus control group.

Ki = net 18F-FDG uptake rate; Kis = specific net 18F-FDG uptake rate.

Fig. 5. Regional net 18F-fluorodeoxyglucose uptake rate (Ki) and specific Ki (Kis) in supine sheep after 2 h of mechanical ventilation with positive end-expiratory pressure = 0 cm H2O and plateau pressure of 30–32 cm H2O, without (control, n = 6, A, B) and with mild endotoxemia (endotoxin, n = 6, C, D). Control values of Ki and Kis (A, B) are quite homogeneous within and between animals. Mild endotoxemia produced an increase in the average whole-lung values of Ki and Kis, with significant regional heterogeneity (C, D), larger in dorsal regions. * P < 0.05 between middle and ventral regions; †P < 0.05 dorsal versus ventral regions; ‡P < 0.05 dorsal versus middle regions; #P < 0.05 endotoxin versus control group. Circles represent individual data points.

Fig. 6. Specific regional net 18F-fluorodeoxyglucose uptake rate (Kis) in supine sheep after 2 h of mechanical ventilation with positive end-expiratory pressure = 0 cm H2O and plateau pressure of 30–32 cm H2O, without (control, n = 6) and with mild endotoxemia (endotoxin, n = 6). Each lung region corresponds to one third of the lungs segmented according to the amount of perfusion (A, B) or gas fraction (C, D). Low, intermediate, and high in the horizontal axes correspond to the first, second, and third tertiles, respectively. Note the linear relationship between perfusion and Kis as opposed to a relationship with a minimum at intermediate values between gas fraction and Kis. * P < 0.05 middle versus ventral regions; †P < 0.05 dorsal versus ventral regions; ‡P < 0.05 dorsal versus middle regions. Circles represent individual data points.
Histologic Findings

Neutrophil counts in the dorsal regions of the endotoxin group were more than double the counts in the same regions of the control group (P < 0.05), whereas no significant difference was present in the ventral regions (table 3). Moderate or severe alveolar hemorrhage was focal and present in only 11% of the high-power fields studied. On average, hemorrhage was mild and more intense in the dorsal regions of the control group (table 3). Perivascular and alveolar edema, septal thickening, and capillary congestion were very mild in both groups, without statistically significant differences.

Discussion

The most important findings of the current study were as follows: (1) mild short-term endotoxemia in the presence of heterogeneous lung aeration and perfusion and large tidal volume mechanical ventilation bounded by clinically acceptable pressure limits produced spatially heterogeneous increases in pulmonary neutrophilic inflammation; (2) these regional increases of neutrophilic inflammation were significantly related to both regional aeration and perfusion and were detectable with PET within 2 h of injury; and (3) in the absence of endotoxemia, pulmonary 18F-FDG uptake was low and uniformly distributed at those settings of mechanical ventilation.

Pulmonary 18F-FDG uptake has been shown to be a marker of the concentration and degree of activation of neutrophils in the nontumoral lung.6,28 We used the model of Sokoloff et al.50 to derive regional parameters from 18F-FDG kinetics. This choice was based on the observation of "Sokoloff-type" tracer kinetics in both control and endotoxin groups (fig. 2), according to a previously described strategy for model selection.30 Based on that analysis, Sokoloff-type kinetics is suggestive of a minimal degree of lung edema.30

The ventilator settings chosen for the study produced in both groups the intended heterogeneity in lung expansion with significant fraction of nonaerated and poorly aerated units in dependent regions (PEEP = 0) and a predominance of normal aeration in nondependent regions (limitation of Pplat to 30–32 cm H2O). In fact, nondependent areas of the control group presented values of Kᵢ and Kᵢₛ that matched those observed in unjured lungs of prone sheep in two previous studies29,30 and did not show any detectable histopathologic injury. PEEP = 0 was the likely cause for the decrease in Fgas in both groups after 2 h of mechanical ventilation. As intended, the used dose of endotoxin did not lead to changes in perfusion distribution to ventral, middle, and dorsal regions (fig. 3B). This result emphasizes the small dose used in our study, which was either equivalent to or lower than doses considered mild and devoid of lung injurious effects in previous investigations.6,13 The nonaerated lung compartment increased in the most dependent regions with endotoxemia. At least three factors could account for this observation: surfactant dysfunction, increase in regional blood volume, and regional edema. All three are compatible with the increased regional shunt found in dependent regions. Given the very mild degrees of edema apparent on histologic samples, the two first factors are the most likely.

Endotoxin has been shown to induce systemic inflammation and to increase sequestration of neutrophils in the lungs yielding increased whole-lung 18F-FDG uptake.4–6 Previous ex vivo and in vivo small animal models described an increase in the release of inflammatory mediators and a modification in inflammatory cell infiltration due to endotoxin, which depended on tidal volume.4,11,12,14 However, no previous study has investigated whether and how those results can be extrapolated to the heterogeneously aerated and perfused large animal lung. Such extrapolation is complex, given that mechanical strain is heterogeneously distributed in the me-
mechanically ventilated heterogeneously aerated lung and that there is still controversy over the contribution of overdistension and low volume ventilation to inflammation during ventilator-induced lung injury. Thus, it is difficult to predict how mild endotoxemia in the presence of heterogeneous pulmonary perfusion would interact with that heterogeneous distribution of lung inflation to affect lung inflammation, a condition related to clinical situations such as intraoperative mechanical ventilation during surgical interventions involving subclinical endotoxemia. A previous large animal study showed that endotoxemia produced increased whole lung neutrophil activation. However, no data were provided on regional inflammation or on the combined effect of endotoxemia and mechanical ventilation. Understanding regional heterogeneities is essential to optimize the use of strategies to reduce lung injury by reducing heterogeneity of aeration and perfusion, according to the degree of lung injury severity.

Our results indicate that the combination of mild endotoxin doses with ventilatory settings, which did not by themselves cause injury, resulted in a substantial increase in pulmonary neutrophilic inflammation. This increase showed a heterogeneous spatial distribution, characterized by a vertical dependence of neutrophilic inflammation (K_i and Ki_s), more intense in dependent regions. The fact that the observed increase in \(^{18}\)F-FDG uptake was still significant after correction for regional lung tissue (K_i), including correction for regional lung collapse and blood volume, supports the conclusion that it was not a mere consequence of the increase in the amount of pulmonary tissue per unit volume of lung or in regional blood volume in derecruited regions because it was still significant after correction for regional lung tissue (K_i). The increase in neutrophil counts in dependent regions of the endotoxin group reinforces the imaging findings.

Remarkably, changes in global and regional \(^{18}\)F-FDG uptake in the endotoxemia group were markedly larger than those in regional aeration and perfusion. The average K_i and K_i_s in the endotoxin group were more than twofold compared with those in the control group, whereas average F_gas and pulmonary perfusion distributions were similar in the two groups. Such similarity supports the inference that perfusion distribution per se could not explain the changes in global and regional \(^{18}\)F-FDG uptake during endotoxemia. Furthermore, although deterioration of regional aeration and shunt in the endotoxin group was limited to the dorsal regions, and distributions of F_gas and perfusion were similar in the two groups, \(^{18}\)F-FDG uptake (K_i) in the endotoxin group was larger than that in the control group in all lung regions: 47% larger in ventral regions, 103% in middle regions, and 150% in dorsal regions. These results suggest that \(^{18}\)F-FDG may be an early and sensitive marker of regional pulmonary inflammation induced by the combination of endotoxemia and ventilator-induced lung injury.

Endotoxemia could contribute to the development of regional inflammation during mechanical ventilation by activation of neutrophils, resulting in their augmented response to the inflammatory stimuli produced by localized mechanical strain, which could be excessive in heterogeneous lungs. Our findings are, therefore, in line with the two-hit theory, although we used both injurious stimuli simultaneously and not sequentially as in typical two-hit studies. The changes in circulating neutrophil counts suggest another form of interaction between mechanical ventilation and endotoxemia. The nonsignificant increase of circulating neutrophils in the control group, a trend noted in previous studies, might be explained by release from the marginalized pool due to mechanical ventilation. The lack of migration of these cells to the lungs or the lack of activation of these cells in the lungs implies that an additional hit would have a greater than normal supply of neutrophils to call on. The observed fall in circulating neutrophils in the endotoxin group shows that this occurred and that neutrophils have either migrated in the capillaries or migrated into the lungs. The increase in K_i and K_i_s in the endotoxin group indicates that these neutrophils became activated.

The multivariate regression analysis showed that both perfusion and regional aeration were important to explain the observed regional values of K_i. The linear relationship between K_i and regional perfusion (fig. 6) could represent the increase in regional inflammation with the increase in regional load of endotoxin, inflammatory cells, including neutrophils, and mediators of inflammation to the more perfused regions predominant in the dependent lung. This association between regional lung perfusion and K_i is important because therapeutic interventions can modify perfusion distribution, for example, as recently shown with noisy ventilation. In contrast to the direct relationship between K_i and regional perfusion, the relationship between K_i and F_gas was biphasic with large K_i in the low and high extremes of aeration and low K_i for F_gas values in the normal range (fig. 6). The large K_i for low F_gas may represent the contribution of low volume lung injury associated with nonaerated and poorly aerated predominantly in dependent regions, whereas the high K_i for larger F_gas values suggest the contribution of lung overdistension. Although we did not find a significant fraction of hyperinflated areas, our measurements performed at mean lung volume underestimate hyperinflation, as discussed below. Thus, we speculate that overdistension could have occurred at end inspiration in regions of high F_gas.

Despite the substantial heterogeneity in lung aeration, we found a low and uniform distribution of neutrophilic inflammation in the control group. Such regional findings contrast with commonly invoked interdependence mechanisms, which predict that the used P_plat would generate high local forces in regions lying in the interface between aerated and derecruited lung. These forces would be expected to result in inflammation and increased \(^{18}\)F-FDG uptake. At least two factors could account for our finding. First, \(^{18}\)F-FDG uptake could be an insensitive marker of regional lung mechanical injury. However, we showed that peak pressures of 50 cm H_2O applied for 90 min increased lung \(^{18}\)F-FDG uptake in homogeneously expanded sheep lungs, supporting the sensi-

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tivity of the technique. Given that interdependence mechanisms would predict local pressures higher than 130 cm H$_2$O for $P_{\text{plat}} = 30–32$ cm H$_2$O, if those local values were present, they should be detectable with our technique. Indeed, our histologic findings support the absence of parenchymal inflammation in the control group. Consequently, the second possibility is that the assumptions of that previously theorized interdependence model may not accurately represent the expansion of the dorsal regions of a normal lung.

Advantages and limitations of the used imaging techniques have been discussed in detail previously. Specifically to this research, the regional aeration of the lung was computed from PET transmission scans. Because these scans are collected for 10 min during uninterrupted mechanical ventilation, the calculated $F_{\text{gas}}$ of a region represents its average aeration over the breathing cycle. $F_{\text{gas}}$ is not only affected by motion but also by filtering during image reconstruction and processing, and partial volume effects. The consequent limited spatial resolution (13 mm) could result in underestimation of the degree of regional hyperinflation compared with that measured from computed tomography images acquired during end-inspiratory breath holds at much higher spatial resolution. Additional limitations include (1) species differences: young sheep as studied in our work are known to have reduced collateral ventilation, which might make them more prone than adult humans to reabsorption atelectasis and lung collapse. Also, presence of pulmonary intravascular macrophages in sheep may make them differently sensitive to endotoxia-induced lung injury when compared with humans. (2) Ventilatory settings: we chose settings aimed at maximizing aeration heterogeneity. Settings used clinically might be associated with increased spatial heterogeneity of inflammation during ALI due to endotoxia and mechanical ventilation. The authors thank Wellington V. Cardoso, M.D., Ph.D. (Professor of Medicine and Pathology, Boston University School of Medicine, Boston, Massachusetts), and Mauro Tucci, M.D., Ph.D. (Research Fellow, Department of Anesthesia and Critical Care, Massachusetts General Hospital and Harvard Medical School, Boston, Massachusetts, and Respiratory Intensive Care Unit, University of Sao Paulo School of Medicine, Sao Paulo, Brazil), for support with histopathologic analysis; Hui Zheng, Ph.D. (Assistant Professor, Biostatistics Center, Massachusetts General Hospital), for assistance with statistical analysis; Steven B. Weise, B.S. (Senior Research Technician, Division of Nuclear Medicine, Massachusetts General Hospital), for preparation of the radioisotopes; and Ivan Schettino, M.D., Ph.D., Tommaso Mauri, M.D., and Maria Avila, M.D. (Research Fellows, Department of Anesthesia and Critical Care, Massachusetts General Hospital and Harvard Medical School), for assistance with animal preparation.

References


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Underwood’s Inspirator

Before extending short-lived corporate branches to Canada and England, New York’s G.B. Underwood Inspirator and Oxygen Company had popularized heating and medicating oxygen for treating an astonishing array of afflictions: anemia, asphyxia, asthma, blood poisoning, bronchitis, catarrh, cholera, consumption, croup, diabetes, diphtheria, hay fever, laryngitis, otitis media, pneumonia, tinnitus, typhoid fever, and even tuberculosis (both pulmonary and laryngeal). Advertisements assured that “oxygen heated by the Underwood 20th Century Pulmonary Inspirator is assimilated three times more readily and plentifully than when given cold.” Noting that “one cylinder heated will do the work of three cold,” G.B. Underwood underscored the economy of the Inspirator for rural physicians, “who have to pay a heavy freight on the oxygen they use in their cases.” (Copyright © the American Society of Anesthesiologists, Inc. This image appears in color in the Anesthesiology Reflections online collection available at www.anesthesiology.org.)

George S. Bause, M.D., M.P.H., Honorary Curator, ASA’s Wood Library-Museum of Anesthesiology, Park Ridge, Illinois, and Clinical Associate Professor, Case Western Reserve University, Cleveland, Ohio. UJYC@aol.com.